

The First Synthesis of Herbicidin B. Stereoselective Construction of the Tricyclic Undecose Moiety by a Conformational Restriction Strategy Using Steric Repulsion between Adjacent Bulky Silyl Protecting Groups on a Pyranose Ring[†]

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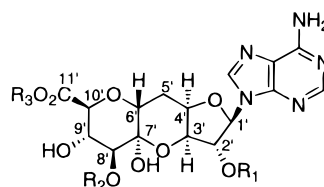
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Abstract: The first total synthesis of the nucleoside antibiotic herbicidin B (**1b**) was achieved, where a novel aldol-type C-glycosidation reaction promoted by samarium diiodide (SmI₂) was used as a key step. Treatment of methyl 3,4-O-(1,1,3,3-tetraisopropyl-1,3-disiloxanediyl)-1-phenylthio-2-ulos-β-D-glucuronate (**13**) with SmI₂ in THF regioselectively gave the corresponding 1-enolate, which was readily trapped with 1-β-D-xylosyladenine 5'-aldehyde derivative **7** to afford the product **19a,b** as an anomeric mixture. Dehydration of the 5'-hydroxyl in **19a,b** with using Burgess's inner salt gave the enone **20**, which was subsequently hydrogenated to give undeculofuranuronyl adenine derivative **21**. Deprotection of **21** gave a tricyclic sugar nucleoside, **23**. However, it was an epimer of herbicidin B at the 6'-position. Construction of the desired 6'-α-configuration was achieved by using a conformational restriction strategy based on repulsion between adjacent bulky protecting groups on the pyranose ring. Thus, when methyl 3-O-tert-butylidimethylsilyl-4-O-tert-butylidiphenylsilyl-1-phenylthio-2-ulos-D-glucuronate (**29c**), the conformation of which was restricted in an unusual ¹C₄-like conformation, was used as a precursor for ulose 1-enolate in the SmI₂-promoted aldol reaction with **7**, the desired 6'-α-aldol product **30c** was predominantly obtained. Compound **30c** was dehydrated, followed by hydrogenation of the alkenyl bond and then deprotection to form an internal ketal linkage between the 3'- and 7'-positions, which spontaneously gave herbicidin B.

Introduction

Nucleoside antibiotics are fascinating compounds that show a variety of biological activities.¹ They are not only used as medicines, lead compounds for the development of drugs, and biological tools but are also important as total synthetic targets.²

Herbicidins (**1a–f**),³ aureonucleomycin (**2**),⁴ and S12245 (**3**),⁵ a series of adenine nucleoside antibiotics that have the same backbone structure (Figure 1), have been isolated from strains of *Streptomyces*. Herbicidin A (**1a**) and B (**1b**), as well as aureonucleomycin (**2**), efficiently inhibit the growth of *Xanthomonas oryzae*, which causes rice leaf blight, and are also selectively toxic toward dicotyledon. These compounds have some interesting structural features: (1) adenine is glycosylated



	R ₁	R ₂	R ₃
Herbicidin A (1a)	Me	CH ₃ CH=C(CH ₂ OH)CO	Me
Herbicidin B (1b)	Me	H	Me
Herbicidin C (1c)	H	H	Me
Herbicidin E (1d)	Me	(CH ₃) ₂ CHCO	Me
Herbicidin F (1e)	Me	CH ₃ CH=C(CH ₃)CO	Me
Herbicidin G (1f)	H	CH ₃ CH=C(CH ₃)CO	H
Aureonucleomycin (2)	Me	CH ₃ CO	Me
S12245 (3)	H	H	H

Figure 1. Structures of herbicidins.

at the 1β-position of an unusual sugar, undecose, which has a tricyclic furano-pyrano-pyran structure; (2) there is an internal hemiketal linkage between the C-3'- and -7'-positions which forms a trans junction for a pyrano-pyran ring; and (3) all of the substituents at the C-7', -8', -9', and -10'-positions on the second pyranose are fixed in axial positions due to the tricyclic structure of the undecose. Although considerable effort has been devoted to the total synthesis of herbicidins because of their unique and complex structures,^{6,7} none of these attempts has yet been successful.

In this paper, we describe the synthesis of herbicidin B (**1b**), in which a novel samarium diiodide (SmI₂)-promoted aldol-

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[†] This paper constitutes Part 190 of Nucleosides and Nucleotides. For part 189 in the series, see: Naka, T.; Nishizono, N.; Minakawa, N.; Matsuda, A. *Tetrahedron Lett.* **1999**, *40*, 6297–6300.

(1) (a) Isono, K. *J. Antibiot.* **1988**, *41*, 1711–1739. (b) Isono, K. *Pharmacol. Ther.* **1991**, *52*, 296–286.

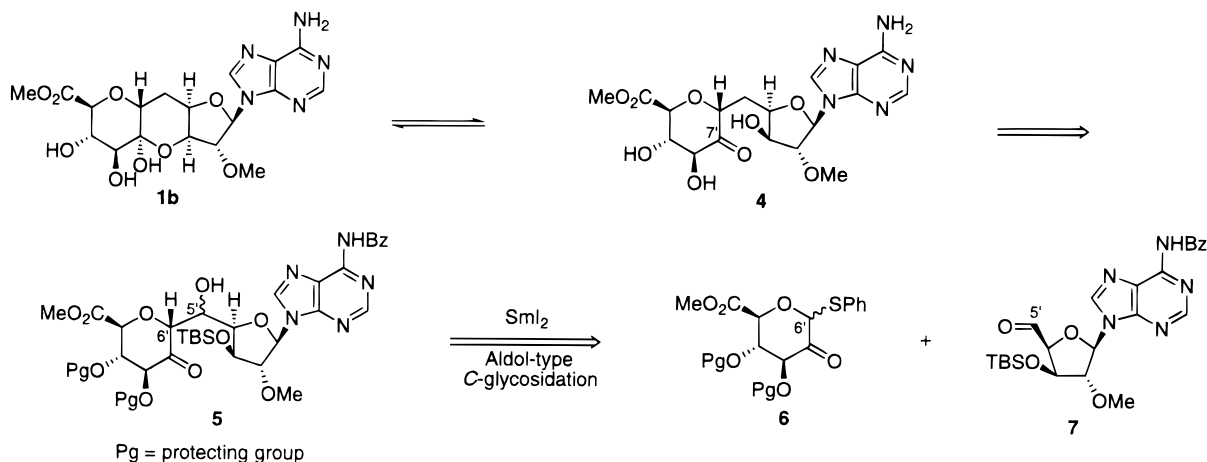
(2) Knapp, S. *Chem. Rev.* **1995**, *95*, 1859–1876.

(3) (a) Arai, M.; Haneishi, T.; Kitahara, N.; Enokita, R.; Kawakubo, K.; Kondo, Y. *J. Antibiot.* **1976**, *29*, 863–869. (b) Haneishi, T.; Terahara, A.; Kayamori, H.; Yabe, J.; Arai, M. *J. Antibiot.* **1976**, *29*, 870–875. (c) Taguchi, Y.; Yoshikawa, H.; Terahara, A.; Torikata, A.; Terao, M. *J. Antibiot.* **1979**, *32*, 857–861. (d) Takiguchi, Y.; Yoshikawa, H.; Terahara, A. *J. Antibiot.* **1979**, *32*, 862–867. (e) Terahara, A.; Haneishi, T.; Arai, M. *J. Antibiot.* **1982**, *35*, 1711–1714.

(4) Dai, X.; Li, G.; Wu, Z.; Lu, D.; Wang, H.; Li, Z.; Zhou, L.; Chen, X.; Chen, W. *Chem. Abstr.* **1989**, *111*, 230661f.

(5) Tsuzuki, M.; Suzuki, G. *Chem. Abstr.* **1988**, *109*, 53206x.

Scheme 1



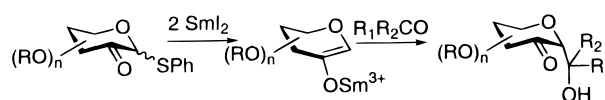
type *C*-glycosidation reaction with 1-phenylthio-2-ulose derivatives, as a precursor for generating ulose-1-enolate,⁸ is effectively used as a key step. This is the first total synthesis of a herbicidin congener.

Results and Discussion

Retrosynthetic Analysis and Strategy. Our retrosynthetic analysis and synthetic strategy for herbicidin B (**1b**) are shown in Scheme 1. Herbicidin B can be considered the equivalent of a ring-opened 7'-keto compound, **4**, and appropriate protection of the hydroxyl groups and the introduction of a hydroxyl group at the C-5' of **4** leads to aldol product **5**. This aldol product **5** is then retrosynthetically disassembled by an aldol-type reaction to the phenylthioulose **6** as a precursor of enolate and 1- β -D-xylosyladenine 5'-aldehyde derivative **7** as an acceptor. Compounds **6** and **7** can be synthesized from D-glucose and adenosine, respectively.

In this retrosynthetic analysis and synthetic strategy, the following two steps are considered key steps: (a) formation of the tricyclic structure with the desired stereochemistry, namely 6'- α -(6',7'-*trans*)-configuration, in the second pyranose ring in which the substituents at the C-8', -9', and -10' positions are all in axial positions, and (b) a stereoselective *C*-glycosidation which gives the 6'- α -configuration by an aldol-type reaction between a ulose derivative and an adenine nucleoside 5'-aldehyde. In the former step, we expected that spontaneous cyclization by the formation of a ketal linkage between the 3'- and 7'-positions would be achieved in a stereoselective manner to give the desired herbicidin B when the protecting groups of the hydroxyls are removed. In the latter step, we planned to use a SmI₂-promoted aldol-type *C*-glycosidation reaction, which we recently developed (Scheme 2).⁸ This reaction is very useful because (1) phenylthio-2-ulose derivatives, the precursor for generating enolates, are stable and easy to prepare, (2) regioselective enolization at the anomeric position of the ulose

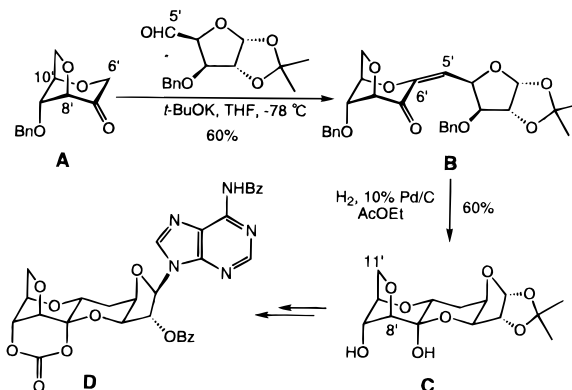
Scheme 2



derivative is possible by SmI₂-promoted reductive cleavage of a C-S bond at the anomeric position,⁹ (3) substrates such as nucleoside 5'-aldehydes, which are unstable under basic and/or acidic conditions,¹⁰ can be used, since the reaction proceeds under neutral conditions at -78 °C, and (4) the reaction gives the corresponding α -*C*-glycosides as a major product in high yields. Based on these findings, we expected that the 6'- α -aldol product **5** could be obtained by the SmI₂-promoted condensation reaction between **6** and **7**.

Preparation of the 1-Phenylthio-2-ulose Unit. We first selected **13** as a precursor for ulose 1-enolate, in which the 3,4-*trans* hydroxyls were protected by a 1,1,3,3-tetraisopropylidisiloxane (TIPDS) group. Scheme 3 shows its preparation. Protection of the 4- and 6-hydroxyls of phenyl 1-thio- β -D-glucoside (**8**) by a TIPDS group, followed by acid-catalyzed

(9) Gallagher et al. reported that the generation of the 1-enolate was successful when 3,6-anhydro-ulose **A** was used as a substrate. Compound **B** was further glycosylated with adenosine to give nucleoside derivative **D**. However, hydrolysis of the 8',11'-ether linkage on **D** was unsuccessful; see ref 7c.

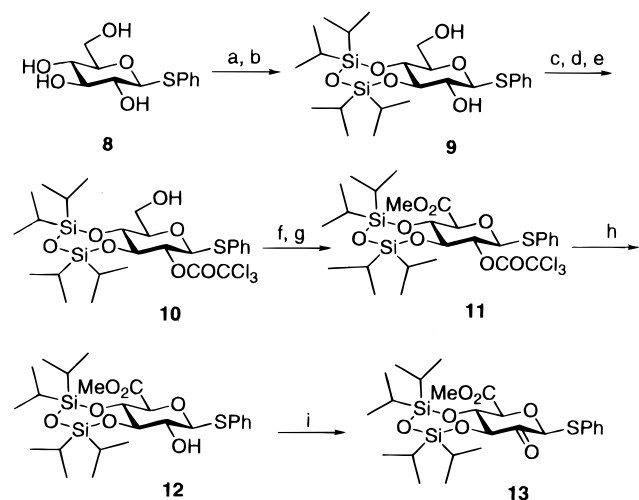


(10) Gallagher et al. tried to prepare an aldol condensation product between **A** (footnote 9) and adenosine 5'-aldehyde derivative which was almost the same as **7**, but this was unsuccessful due to the instability of adenosine 5'-aldehyde under the basic conditions used. Therefore, reductive coupling using ulosyl bromides as enolates was developed. However, ulosyl bromides are not very stable, and the substrates that can be synthesized are limited: Binch, H. M.; Griffin, A. M.; Schwidetzky, S.; Ramsay, M. V. J.; Gallagher, T.; Lichtenthaler, F. W. *J. Chem. Soc., Chem. Commun.* **1995**, 967-968.

(6) (a) Beader, J. R.; Dewis, M. L.; Whiting, D. A. *J. Chem. Soc., Perkin Trans. 1* **1995**, 227-233. (b) Fairbanks, A. J.; Perrin, E.; Sinay, P. *Synlett* **1996**, 679-681. (c) Emery, F.; Vogel, P. *J. Org. Chem.* **1995**, *60*, 5843-5854. (d) Emery, F.; Vogel, P. *Tetrahedron Lett.* **1994**, *34*, 4209-4212.

(7) (a) Binch, H. M.; Griffin, A. M.; Gallagher, T. *Pure Appl. Chem.* **1996**, *68*, 589-592. (b) Newcombe, N. J.; Mahon, M. F.; Molloy, K. C.; Alker, D.; Gallagher, T. *J. Am. Chem. Soc.* **1993**, *115*, 6430-6431. (c) Binch, H. M.; Gallagher, T. *J. Chem. Soc., Perkin Trans. 1* **1996**, 401-402.

(8) Ichikawa, S.; Shuto, S.; Matsuda, A. *Tetrahedron Lett.* **1998**, *39*, 4525-4528. SmI₂-promoted *C*-glycosidation reaction with mannosyl pyridylsulfones as substrates were also reported: Jarretton, O.; Skrydstrup, T.; Beau J.-M. *Tetrahedron Lett.* **1997**, *38*, 1767-1770.

Scheme 3^a

^a Reagents: (a) TIPDSCl₂, pyridine; (b) *p*-TsOH, DMF; (c) TrCl, pyridine; (d) Cl₃COCl; (e) TFA, CH₂Cl₂; (f) DCC, TFA, DMSO; (g) PDC, MeOH, DMF; (h) K₂CO₃, MeOH; (i) (COCl)₂, DMSO, Et₃N, CH₂Cl₂.

isomerization,¹¹ gave **9** in 59% yield in two steps. After protection of the 2-hydroxyl of **9** by a trichloroacetyl group, it was successively treated by Moffatt oxidation conditions and PDC in the presence of MeOH¹² to give methyl ester derivative **11** in 61% yield. Removal of the trichloroacetyl group at the 2-position of **11** with K₂CO₃/MeOH, followed by Swern oxidation of the resulting secondary alcohol, gave 1-phenylthio-2-ulose derivative **13** in excellent yield. The compound **13** was sufficiently stable to be purified by silica gel column chromatography.

Preparation of the 1-β-D-Xylosyladenine 5'-Aldehyde Unit.

1-β-D-Xylosyladenine 5'-aldehyde derivative **7** was prepared as shown in Scheme 4. After the 5'-primary hydroxyl of 2'-*O*-methyladenosine **14** (prepared by a known method¹³) was protected by a triphenylmethyl (Tr) group, the 3'-hydroxyl was inverted by a successive oxidation–reduction procedure with CrO₃/Ac₂O/pyridine and NaBH₄/AcOH systems¹⁴ to give xylo-nucleoside **16** in 78% yield. Protection of the resulting 3'-hydroxyl and 6-amino groups of **16** with *tert*-butyldimethylsilyl (TBS) and benzoyl groups, respectively, by usual procedures gave **17** in 83% yield. Removal of the Tr group of **17** under acidic conditions, followed by oxidation with Dess–Martin periodinane, gave **7**, an acceptor of the aldol reaction. The aldehyde **7** was used for the next reaction without further purification because of its instability.

Synthesis of 6'-epi-Herbicidin B Using a SmI₂-Promoted Aldol-Type C-Glycosidation Reaction as a Key Step.

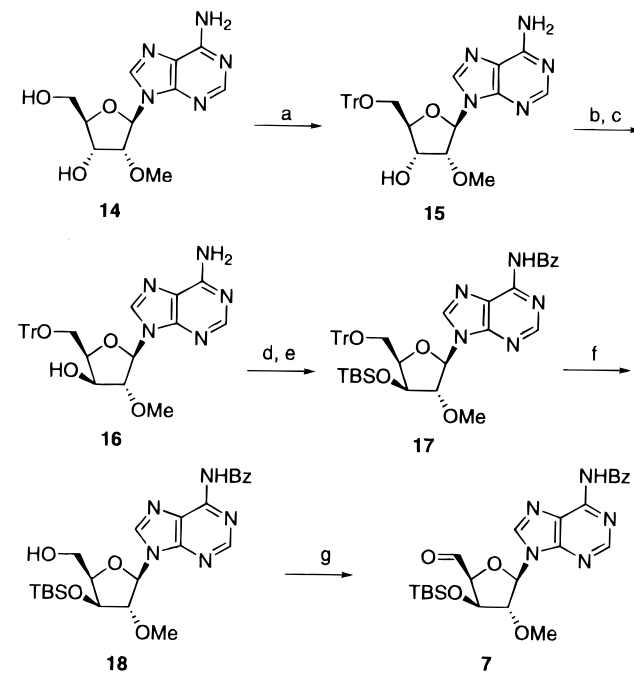
We examined the SmI₂-promoted aldol-type C-glycosidation reaction with **13** and **7** (Scheme 5). When a solution of **13** in THF was added dropwise to a stirring solution of SmI₂ (2.0 equiv) in THF at –78 °C, the corresponding samarium I-enolate was successfully generated, to which a solution of **7** (1.0 equiv) in THF was added to give the desired aldol products in 75% yield as an anomeric mixture (**19a**/**19b** = 79/21). NOE experiments of the mixture revealed their 6'-stereochemistries; correlation

(11) van Boeckel, C. A. A.; van Boom, J. H. *Tetrahedron* **1985**, *41*, 4545–4555.

(12) Garegg, P. J.; Olsson, L.; Oscarson, S. *J. Org. Chem.* **1995**, *60*, 2200–2204.

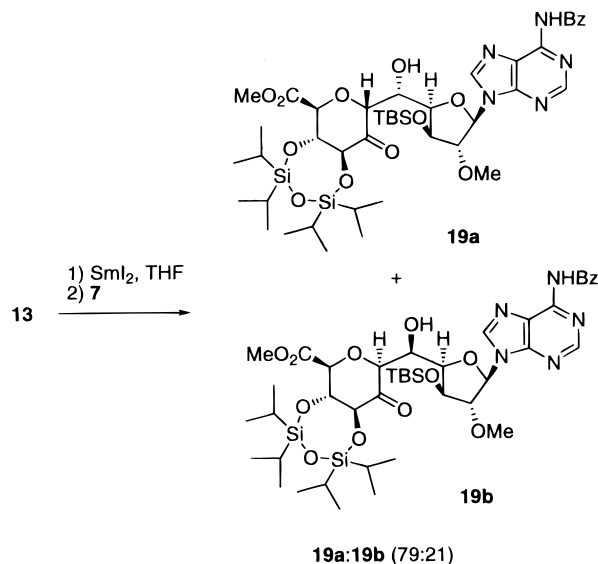
(13) Yano, J.; Kan, L. S.; Ts'o, P. O. P. *Biochim. Biophys. Acta* **1980**, *629*, 178–183.

(14) Robins, M. J.; Samano, V.; Johnson, M. D. *J. Org. Chem.* **1990**, *55*, 410–412.

Scheme 4^a

^a Reagents: (a) TrCl, pyridine; (b) CrO₃, pyridine, Ac₂O, 4-Å molecular sieves, CH₂Cl₂; (c) NaBH₄, AcOH; (d) TBSCl, imidazole; (e) BzCl, pyridine; (f) TFA, CH₂Cl₂; (g) Dess–Martin periodinane, CH₂Cl₂.

Scheme 5



between H-6' and H-9' for the α-anomer **19a** and correlations between H-6' and H-8', and between H-6' and H-10', for the β-anomer **19b** were observed (Figure 2).¹⁵

Radical deoxygenation of the 5'-hydroxyl group of the anomeric mixture **19a,b** via the corresponding 5'-*O*-methyl oxalate¹⁶ or 5'-thiocarbonylimidazolide¹⁷ was tried but was

(15) The 5'-configurations of **19a** and **19b** were suggested to be 5'*R* and 5'*S*, respectively, on the basis of the possible six-membered-ring chelation transition state in the SmI₂-mediated aldol reaction. In fact, the 5'*R*-configuration of **19a** was confirmed as shown below. During the treatment of **19a** with SmI₂ in MeOH, an isomerization of the configuration at the 6'-position was observed.

(16) (a) Matsuda, A.; Takenuki, K.; Itoh, H.; Sasaki, T.; Ueda, T. *Chem. Pharm. Bull.* **1987**, *35*, 3967–3970. (b) Matsuda, A.; Takenuki, K.; Sasaki, T.; Ueda, T. *J. Med. Chem.* **1991**, *34*, 235–239. (c) Dolan, S. C.; MacMillan, J. J. *Chem. Soc., Chem. Commun.* **1985**, 1588–1589.

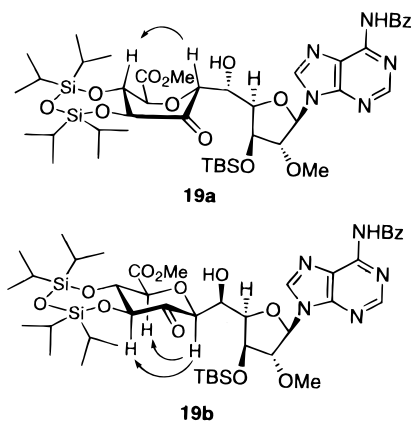
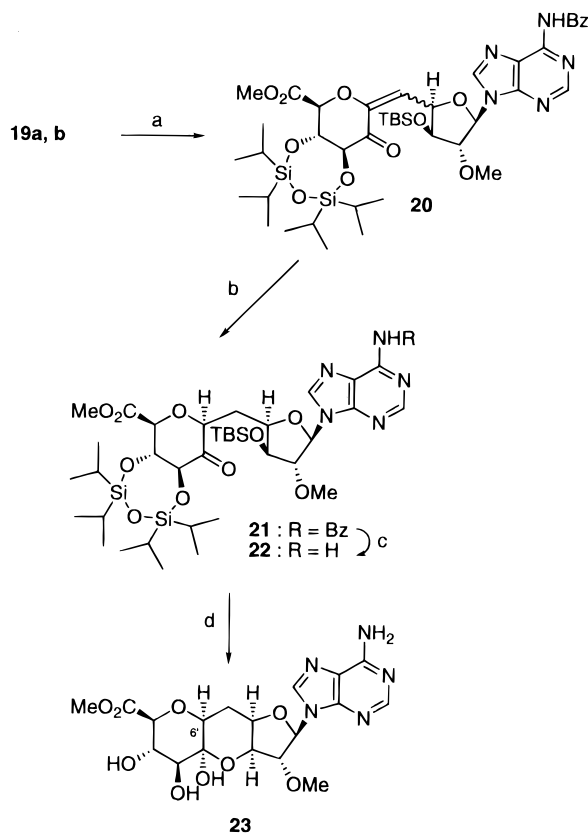


Figure 2. Determination of stereochemistry at the 6'-position of **19a** and **19b** by NOE experiments.

Scheme 6^a



^a Reagents: (a) MeO₂CNSO₂NEt₃, toluene; (b) H₂, Pd/C, EtOAc; (c) Sm, I₂, MeOH; (d) TBAF, THF.

unsuccessful.¹⁸ We next sought to dehydrate it by elimination of the 5'-hydroxy of **19a,b**, followed by hydrogenation of the resulting alkenyl bond. Thus, **19a,b** was converted to the corresponding 5'-*O*-mesylate and -triflate. However, treatment of these 5'-*O*-sulfonates with a base led only to their decomposition, and the desired elimination product **20** was not obtained. On the other hand, as shown in Scheme 6, when **19a,b** was treated with MeO₂CNSO₂NEt₃ (Burgess's inner salt)¹⁹ in toluene, the desired elimination reaction proceeded to give the

(17) Pankiewicz, K.; Matsuda, A.; Watanabe, K. A. *J. Org. Chem.* **1982**, *47*, 485–488.

(18) It has been reported that deoxygenation of the 5-hydroxyl group in undecose derivatives was very difficult, possibly because of the instability of the aldol products and significant steric hindrance around the hydroxyl: Cox, P. J.; Griffin, A. M.; Newcombe, N. J.; Lister, S.; Ramsay, M. V. J.; Alker, D.; Gallagher, Y. *J. Chem. Soc., Perkin Trans. 1* **1994**, 1443–1447.

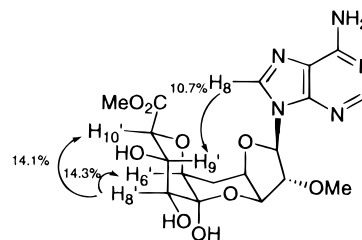


Figure 3. Structure determination of 6'-*epi*-herbicidin B (**23**) by NOE experiments.

enone **20** in 49% yield as a mixture of inseparable isomers due to a *cis/trans* geometry at the 5'-position. The enone **20** was hydrogenated with H₂ in the presence of palladium-on-carbon in EtOAc to give **21** quantitatively, the anomeric configuration of which was determined to be the undesired β by NOE experiments; correlations between H-6' and H-8', and between H-6' and H-10', were observed.²⁰ However, we expected that isomerization at the 6'-position, which was adjacent to the 7'-carbonyl, into the desired α -configuration might occur in the following stages.

Therefore, removal of the protecting groups was examined next. When **21** was treated with NaOMe or K₂CO₃ in MeOH to remove the benzoyl group at the N⁶-position, it decomposed and did not give any of the debenzoylated **22**. However, exposure of **21** to samarium and I₂ in MeOH²¹ successfully gave the debenzoylated product **22**. When **22** was treated with tetrabutylammonium fluoride (TBAF) in THF, all of the silyl protecting groups were removed simultaneously to give a tricyclic sugar product by spontaneously forming an internal ketal linkage between the 3'- and 7'-positions, as expected. However, the product was not herbicidin B, but rather 6'-*epi*-herbicidin B (**23**), an epimer of herbicidin B at the 6'-position. The structure of **23** was confirmed by ¹H NMR and NOE experiments. As shown in Figure 3, the coupling constants, $J_{8,9'} = J_{9',10'} = 9.5$ Hz, suggested its ⁹C_{6'}-structure, and NOE correlations between H-6' and H-8', H-8' and H-10', and H-9' and H-8 at the adenine moiety suggested a β -configuration at the 6'-position. Although isomerization of **23** into herbicidin B was investigated by treating it under acidic and basic conditions, such isomerization was not observed.²²

Conformation-Flip Strategy. Based on the above results, we recognized that it would be essential to provide the 6'- α -*C*-glycosidic structure before forming the 3',7'-ketal linkage to construct the desired tricyclic structure of the sugar moiety of herbicidin B. Therefore, stereoselective hydrogenation from the α -face of the 5',6'-alkenyl bond in the enone system was required. The ¹H NMR spectrum of **20** showed that the substituents at 8', 9', and 10' were in equatorial positions; both

(19) Burgess, E. M.; Penton, H. R.; Taylor, E. A. *J. Org. Chem.* **1973**, *38*, 26–31.

(20) ¹H NMR analysis of the reaction mixture suggested that the hydrogenation product was a mixture of anomers, and the α -anomer readily isomerized during the workup to give the β -anomer as the sole product.

(21) Yanada, R.; Negoro, N.; Bessho, K.; Yanada, K. *Synlett* **1995**, 1261–1263.

(22) Molecular modeling of both herbicidin B (**1b**) and its 6'-epimer **23** was carried out using Macromodel (version 4.5) software. Preferred conformations were determined from the results of Monte Carlo conformer searches starting from previously minimized structures of the two compounds. Minimization employed the MM2* force field. Sets of conformers close to minimum were found for **1b** and **23**, respectively. The minimum-energy conformation for **23** was consistent with that suggested from the coupling constants and NOE relationships of **23**. The minimum-energy conformer of 6'-*epi*-herbicidin B (**23**) was approximately 3.0 kcal/mol more stable than that of herbicidin B.

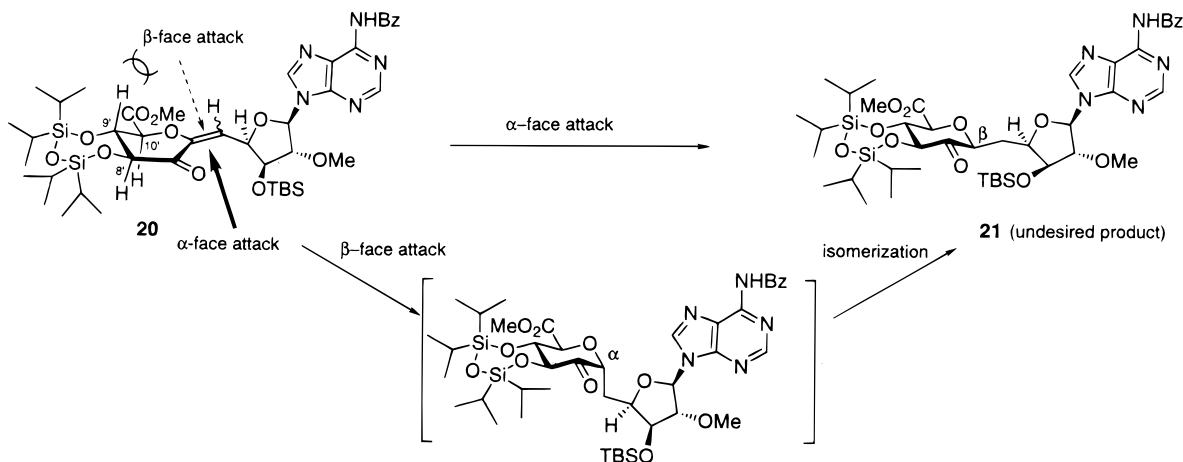


Figure 4.

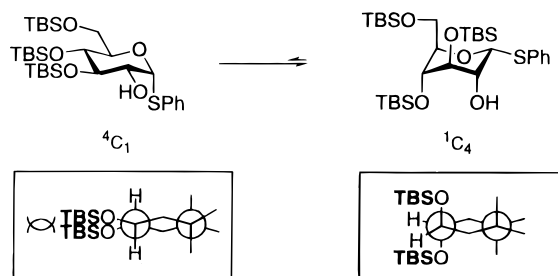


Figure 5.

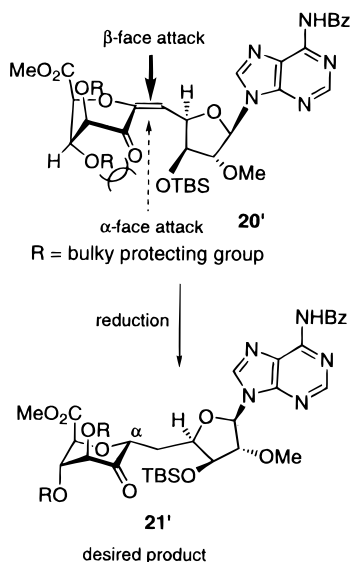
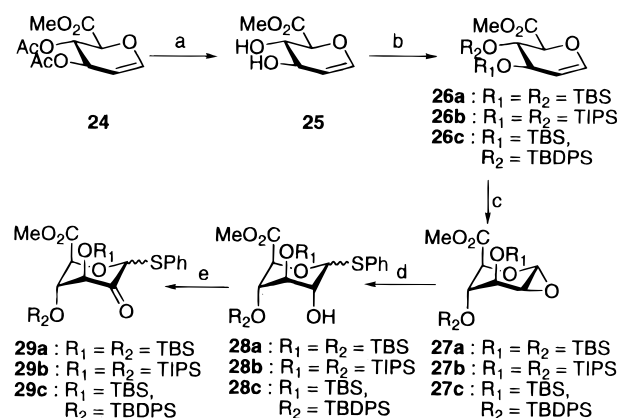


Figure 6.

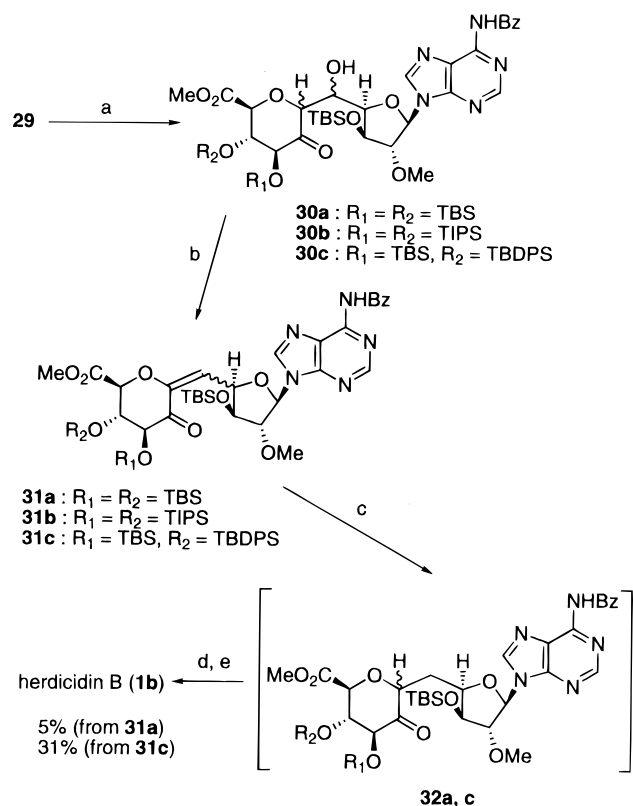
$J_{8,9}$ and $J_{9,10}$ were ca. 9–10 Hz. Accordingly, **20** preferentially adopts a half-chair conformation, as shown in Figure 4. Since hydrogenation from the β -face of the alkenyl bond of **20** would be disfavored probably due to steric repulsion for the 9'-axial proton, α -face attack proceeded to give the undesired 6'- β -C-glycosidic product **21** as the major product. It is also possible that isomerization at the anomeric 6'-position into the undesired β -configuration might occur if the hydrogenation product was the desired α -C-glycoside. ^1H NMR analysis of 6'- β -C-glycoside **21** suggested that it adopts a $^9\text{C}_6$ -like conformation in which all of the substituents on the ulose moiety are in equatorial positions. 6'- β -C-Glycoside **21** would be more thermodynamically stable than the corresponding α -C-glycoside, which has a very bulky 6'-substituent in an axial position, and therefore the undesired isomerization might occur.

Scheme 7^a

^a Reagents: (a) K_2CO_3 , MeOH; (b) TBSCl, TIPSCl, or TBDSPCl, imidazole, DMF; (c) dimethylthioxirane, acetone; (d) PhSH, $\text{BF}_3\text{Et}_2\text{O}$, CH_2Cl_2 ; (e) Dess–Martin periodinane, CH_2Cl_2 .

Recently, it was reported that introducing significantly bulky protecting groups at vicinal hydroxyl groups on pyranoses caused a flip of their conformation from the usual $^4\text{C}_1$ -form into an unusual $^1\text{C}_4$ -form, where the substituents were in axial orientations due to steric repulsion between the bulky protecting groups (Figure 5).²³ We expected that enone **20'**, which has bulky protecting groups at both the 8'- and 9'-hydroxyls, would adopt a $^6\text{C}_9$ -like conformation, as shown in Figure 6, due to steric repulsion between the bulky protecting groups. If so, hydrogenation of **20'** would selectively occur from the β -face to give the desired α -product **21'**, due to a significant steric repulsion for the 9'-axial protecting group when the catalyst accesses the alkenyl bond from the α -face. In addition, the desired α -glycoside **21'** would be more thermodynamically stable than the corresponding β -C-glycoside, since the substituent at the 6'-position of **21'** is in an equatorial position. Thus, isomerization from a β - to an α -configuration at the 6'-position would occur if the hydrogenation does not proceed selectively from the β -face. Based on these considerations, we designed phenylthioglucosides **29a–c** (Scheme 7), which had bulky silyl protecting groups at both the 8'- and 9'-positions, as alternative substrates for the aldol reaction.

(23) (a) Walford, C.; Jackson, R. F. W.; Rees, N. H.; Clegg, W.; Heath, S. L. *Chem. Commun.* **1997**, 1855–1856. As for reactions: (b) Hosoya, T.; Ohashi, Y.; Matsumoto, T.; Suzuki, K. *Tetrahedron Lett.* **1996**, *37*, 663–666. (c) Roush, W. R.; Sebesta, S. P.; Bennett, C. E. *Tetrahedron* **1997**, *53*, 8825–8836. (d) Roush, W. R.; Sebesta, D. P.; James, R. A. *Tetrahedron* **1997**, *53*, 8837–8852.

Scheme 8^a

^a Reagents: (a) SmI₂, THF, then **7**; (b) MeO₂CNSO₂NEt₃, toluene; (c) Pd/C, HCO₂NH₄, MeOH; (d) Sm, I₂, MeOH; (e) TBAF, THF.

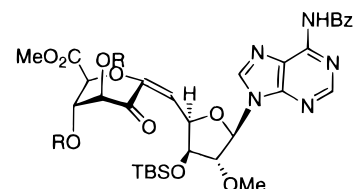
Preparation of Conformation-Flipped 1-Phenylthio-2-ulose Units. The preparation of 1-phenylthio-2-uloses **29a–c** is summarized in Scheme 7. Glycal **24**, derived from D-glucurono-3,6-lactone, was treated with NaOMe in MeOH to give **25**.²⁴ The 3,4-diol of **25** was silylated by usual methods to afford **26a** and **26b**, which have TBS or TIPS groups at both the 3- and 4-hydroxyls, respectively. Glycal **26c** was prepared by successive treatment of **25** with TBSCl and TBDPSCI in the presence of imidazole in DMF. Epoxidation of **26a** with dimethyldioxirane²⁵ gave the gluco-type epoxide **27a** selectively in quantitative yield.²⁶ Next, ring-opening of the epoxide was examined. When **27a** was treated with PhSH and Et₃N, only a trace of the desired phenylthioglycoside **28a** was obtained. Treatment with 10 equiv of PhSH in the presence of a catalytic amount of BF₃·OEt₂ gave **28a** in good yield as an anomeric mixture. Oxidation of **28a** with Dess–Martin periodinane²⁷ gave an anomeric mixture of 1-phenylthio-2-ulose (**29a**) in 80% yield. Similarly, other aldol substrates **29b** and **29c** were prepared from **26b** and **26c**, respectively. The conformations of these phenylthiuloses **29a–c** were investigated by their ¹H NMR spectra. The large coupling constants ($J_{3,4} = 5.8$, $J_{4,5} = 12.8$ Hz) of **29a** showed that it adopted an undesired ⁴C₁-like conformation, similar to that of 3,4-*O*-TIPDS-phenylthiulose **13**. This result suggested that a TBS group would be not bulky enough to flip the conformation. On the other hand, a preference for the desired ¹C₄ conformations in **29b** and **29c**, where the substituents at the 3- and 4-positions are in axial positions, was suggested on

(24) Nishimura, S.; Nomura, S.; Yamada, K. *Chem. Commun.* **1998**, 617–618.

(25) Adam, W.; Chan, Y. Y.; Cremer, D.; Gauss, J.; Scheutzw, D.; Schindler, M. *J. Org. Chem.* **1987**, 52, 2800–2803.

(26) Friesen, R.; Danishefsky, S. J. *J. Am. Chem. Soc.* **1989**, 111, 6656–6660.

(27) Dess, D. B.; Martin, J. C. *J. Am. Chem. Soc.* **191**, 113, 7277–7279.



31a : $J_{8',9'} = 1.7$ Hz, $J_{9',10'} = 3.7$ Hz

31b : $J_{8',9'} = 0$ Hz, $J_{9',10'} = 3.6$ Hz

31c : $J_{8',9'} = 0$ Hz, $J_{9',10'} = 3.3$ Hz

Figure 7. Conformation of the pyranose ring on the enones.

the basis of their coupling constants between H-3 and H-4, and between H-4 and H-5 ($J_{3,4}$, $J_{4,5} = 0–4.3$ Hz).

Synthesis of Herbicidin B. The total synthesis of herbicidin B was achieved as shown in Scheme 8. SmI₂-promoted aldol reactions⁸ with phenylthiulose **29a–c** and aldehyde **7** were examined. When **29a** was treated with SmI₂ at -78 °C, the samarium enolate was not generated efficiently,²⁸ and the yield of the coupling product was low. However, the condensation reaction proceeded effectively at -40 °C, and the desired coupling product **30a** was obtained in 65% yield as a mixture of two diastereoisomers.²⁹ By similar condensation reactions, the aldol products **30b** and **30c** were obtained from **29b** and **29c**, respectively. Treatment of **30a–c** with Burgess's inner salt gave the corresponding enones **31a–c** in 66–79% yields as a single isomer, respectively, while the alkene geometry was unknown.

The conformations of enones **31a–c** were investigated by their ¹H NMR spectra. Small coupling constants between the protons on the pyranose ring, $J_{8',9'} = 0–2$ Hz and $J_{9',10'} = 3–4$ Hz, suggested that they had flipped conformations where the 8'- and 9'-*O*-silyl substituents were in axial positions (Figure 7). Reduction of the enones under a variety of conditions was investigated next. Enone **31b**, which has a pair of significantly bulky TIPS groups at the 8'- and 9'-hydroxyls, was resistant to all of the reduction conditions examined.³⁰ However, when enone **31a**, which has less bulky TBS groups instead of TIPS groups, was treated with HCO₂NH₄ as a hydrogen donor in the presence of Pd–C in MeOH,³¹ the desired reduced product **32a** was obtained.^{32,33} Compound **32a** was then successively treated with SmI₂/MeOH and TBAF/TFH to form an internal ketal linkage between the 3'- and 7'-positions, to spontaneously give a tricyclic sugar compound, the ¹H NMR spectrum of which was consistent with that of herbicidin B (**1b**), while the yield was low (5% from **31a**).

The yield was improved when **31c**, which had a TBS group and a TBDPS group at the 8'- and 9'-hydroxyls, respectively, was used as a substrate. Thus, treatments of **31c** with HCO₂NH₄/Pd–C/MeOH,³³ SmI₂/MeOH, and TBAF/THF gave herbicidin B (**1b**) in 31% yield after preparative silica gel TLC purification. In this reaction, production of 6'-*epi*-herbicidin B (**23**) was not observed at all. This completes the first synthesis of herbicidin B. Its spectroscopic and analytical data are in full agreement with those of the authentic sample.

(28) Remaining starting material **29a** was observed on TLC.

(29) The stereochemistries of the two diastereoisomers were not confirmed.

(30) For example, catalytic hydrogenations with Pd–C, PtO, Raney–Ni, or Pd-black and hydride reductions with DIBAL/HMPA, NaBH₄/NiCl₂, or Bu₃SnH were tried.

(31) Bieg, T.; Szeja, W. *Synthesis* **1985**, 76–77.

(32) The ¹H NMR spectrum of the crude reduction product suggested that it was an anomeric mixture ($\alpha/\beta = 3/1$).

(33) A byproduct, the structure of which has not been determined, was also obtained in the hydrogenation of **31a**, and it made the yield of **32a** low. On the other hand, production of such a byproduct was not observed in the hydrogenation of **31c**.

Conclusions

Using a novel aldol-type *C*-glycosidation reaction promoted by SmI_2 as a key step and a strategy involving the axial orientation of vicinal substituents, we completed the first total synthesis of the nucleoside antibiotic herbicidin B (**1b**). This aldol-type *C*-glycosidation reaction promoted by SmI_2 was very effective for constructing the undecose nucleoside **19** and **30** using 1- β -D-xylosyladenine 5'-aldehyde derivative as an acceptor, which was very unstable under either acidic or basic conditions. The fact that the reactions promoted by SmI_2 proceeded under mild and neutral conditions made this result possible, and since phenylthiouloses as the enolate source are readily synthesized and stable, this approach can be applied to a variety of compounds. Thus, the aldol-type *C*-glycosidation reaction promoted by SmI_2 may be a powerful tool for synthesizing other higher carbon sugar nucleosides. Reduction followed by the deprotection of **20**, in which the conformation of the pyranose ring was 9C_6 , gave selectively 6'-*epi*-herbicidin B, which is an analogue of herbicidin B. On the other hand, similar conversion of enones **31**, which adopted a ${}^{0,8}B$ -type conformation on the pyranose ring, by the introduction of bulky protecting groups gave herbicidin B (**1b**). This work is the first total synthesis of herbicidin congeners, and our methods should be applicable to the synthesis of other herbicidin congeners and their analogues.

Experimental Section

9-(2-O-Methyl-5-O-trityl- β -D-ribofuranosyl)adenine (15). A mixture of **14** (12.9 g, 40 mmol) and TrCl (11.2 g, 44 mmol) in pyridine (100 mL) was stirred for 12 h at 60 °C. After MeOH (10 mL) was added, the solution was evaporated under reduced pressure, and the residue was partitioned between CHCl_3 (500 mL) and H_2O (200 mL). The organic layer was washed with brine (200 mL), dried (Na_2SO_4), and evaporated under reduced pressure. The residue was purified by column chromatography (SiO_2 , 6% MeOH/ CHCl_3) to give **15** (19.2 g, 92% as a white foam): $[\alpha]_D^{+0.29}$ (*c* 1.14, CHCl_3); ${}^1\text{H}$ NMR (CDCl_3 , 500 MHz) δ 8.30 (s, 1H), 8.02 (s, 1H), 7.45–7.23 (m, 15H), 6.14 (d, 1H, $J = 3.7$ Hz), 5.59 (br s, 2H, exchanged with D_2O), 4.50 (dd, 1H, $J = 5.5, 11.1$ Hz), 4.42 (dd, 1H, $J = 3.8, 5.3$ Hz), 4.20 (ddd, 1H, $J = 3.3, 4.5, 11.1$ Hz), 3.57 (s, 3H), 3.52 (dd, 1H, $J = 3.3, 10.7$ Hz), 3.43 (dd, 1H, $J = 4.5, 10.7$ Hz), 2.74 (d, 1H, $J = 6.3$ Hz, exchanged with D_2O); ${}^{13}\text{C}$ NMR (CDCl_3 , 125 MHz) δ 155.8, 153.4, 149.8, 143.8, 139.2, 128.9, 128.8, 128.8, 128.2, 127.5, 120.3, 87.4, 86.9, 84.0, 83.6, 77.5, 77.3, 77.0, 70.0, 63.5, 59.0, 58.5; MS (EI) m/z 523 (M^+). Anal. Calcd for $\text{C}_{30}\text{H}_{29}\text{N}_5\text{O}_4 \cdot 1.2\text{H}_2\text{O}$: C, 66.09; H, 5.80; N, 12.85. Found: C, 66.03; H, 5.56; N, 12.64.

9-(2-O-Methyl-5-O-trityl- β -D-xylofuranosyl)adenine (16). To a mixture of CrO_3 (11.2 g, 112 mmol) and 4-Å molecular sieves (28 g) in CH_2Cl_2 (250 mL) was added pyridine (18.0 mL, 224 mmol) at 0 °C. The mixture was stirred for 30 min at 0 °C, and then Ac_2O (10.6 mL, 112 mmol) was added at the same temperature, and the whole was stirred for additional 15 min. To the resulting mixture was added **15** (14.6 g, 27.9 mmol) in CH_2Cl_2 (100 mL), and the mixture was stirred for 30 min at room temperature. The solution was diluted with EtOAc (1 L) and filtered through a Celite pad, and the filtrate was concentrated to about 500 mL under reduced pressure. The solution was washed with H_2O (3×200 mL) and brine (200 mL), dried (Na_2SO_4), and evaporated under reduced pressure. A mixture of the residue and NaBH_4 (6.87 g, 181 mmol) in AcOH (300 mL) was stirred for 12 h at 4 °C. The mixture was evaporated under reduced pressure, and the residue was coevaporated with EtOH (3×30 mL). The residue was partitioned between CHCl_3 (500 mL) and H_2O (2×200 mL), and the organic layer was washed with saturated NaHCO_3 (200 mL) and brine (200 mL), dried (Na_2SO_4), and evaporated under reduced pressure. The residue was crystallized from EtOAc to give **16** (11.3 g, 78%): mp 205–206 °C; $[\alpha]_D^{+38.8}$ (*c* 0.82, CHCl_3); ${}^1\text{H}$ NMR (CDCl_3 , 500 MHz) δ 8.23 (s, 1H), 7.92 (s, 1H), 7.44 (d, 6H, $J = 7.4$ Hz), 7.26–7.18 (m,

9H), 6.86 (d, 1H, $J = 10.6$ Hz), 5.80 (br s, 2H, exchanged with D_2O), 5.77 (d, 1H, $J = 1.5$ Hz), 4.27 (dd, 1H, $J = 5.6, 8.7$ Hz), 4.21 (dd, 1H, $J = 3.0, 10.5$ Hz), 4.18 (d, 1H, $J = 1.5$ Hz), 3.45 (s, 3H); ${}^{13}\text{C}$ NMR (CDCl_3 , 125 MHz) δ 152.6, 144.0, 140.9, 128.9, 127.9, 127.2, 127.1, 126.7, 91.9, 90.9, 87.3, 82.9, 73.9, 62.4, 58.3; MS (EI) m/z 523 (M^+). Anal. Calcd for $\text{C}_{30}\text{H}_{29}\text{N}_5\text{O}_4$: C, 68.82; H, 5.58; N, 13.38. Found: C, 68.61; H, 5.74; N, 13.19.

N^6 -Benzoyl-9-(3-O-*tert*-butyldimethylsilyl-2-O-methyl-5-O-trityl- β -D-xylofuranosyl)adenine (17). A mixture of **16** (1.48 g, 2.82 mmol), TBSCl (852 mg, 5.62 mmol), and imidazole (767 mg, 11.3 mmol) in DMF (30 mL) was stirred for 15 h at room temperature. After MeOH (5 mL) was added, the mixture was stirred for an additional 12 h and evaporated under reduced pressure. The residue was partitioned between EtOAc (200 mL) and H_2O (100 mL), and the organic layer was washed with brine (50 mL), dried (Na_2SO_4), and evaporated under reduced pressure. A mixture of the residue and BzCl (0.72 mL, 6.2 mmol) in pyridine (30 mL) was stirred for 3 h at room temperature. After NH_4OH (28%, 10 mL) was added to the mixture, the resulting mixture was stirred for a further 20 min and evaporated under reduced pressure. The residue was partitioned between EtOAc (200 mL) and H_2O (100 mL), and the organic layer was washed with brine (50 mL), dried (Na_2SO_4), and evaporated under reduced pressure. The residue was purified by column chromatography (SiO_2 , 33% EtOAc/hexane) to give **17** (1.73 g, 83% as a white foam): $[\alpha]_D^{-27.3}$ (*c* 1.10, CHCl_3); ${}^1\text{H}$ NMR (CDCl_3 , 500 MHz) δ 9.23 (br s, 1H, exchanged with D_2O), 9.00 (s, 1H), 8.30 (s, 1H), 8.19 (d, 2H, $J = 7.5$ Hz), 7.98 (d, 1H, $J = 7.5$ Hz), 7.66–7.40 (m, 17H), 6.50 (s, 1H), 4.81 (dd, 1H, $J = 3.3, 3.7$ Hz), 4.36 (s, 1H), 4.09 (s, 1H), 3.81 (dd, 1H, $J = 3.7, 10.5$ Hz), 3.80 (s, 3H), 3.43 (dd, 1H, $J = 3.3, 10.3$ Hz), 0.73 (s, 9H), –0.76 (s, 3H), –0.89 (s, 3H); ${}^{13}\text{C}$ NMR (CDCl_3 , 125 MHz) δ 152.9, 144.0, 141.9, 132.9, 132.2, 129.1, 128.9, 128.8, 128.1, 128.0, 127.6, 127.4, 90.8, 88.9, 87.4, 84.3, 63.5, 58.6, 25.6, 18.0, –4.9, –5.3; MS (EI) m/z 498 ($\text{M}^+ - \text{Tr}$). Anal. Calcd for $\text{C}_{43}\text{H}_{47}\text{N}_5\text{O}_5\text{Si} \cdot 0.5\text{H}_2\text{O}$: C, 68.77; H, 6.44; N, 9.33. Found: C, 69.06; H, 6.43; N, 9.37.

N^6 -Benzoyl-9-(3-O-*tert*-butyldimethylsilyl-2-O-methyl- β -D-xylofuranosyl)adenine (18). A mixture of **17** (7.31 g, 9.86 mmol) and 80% aqueous TFA (5 mL) in CHCl_3 (100 mL) was stirred for 1 h at room temperature. The mixture was washed with H_2O (2×50 mL), saturated NaHCO_3 (50 mL), and brine (30 mL), dried (Na_2SO_4), and evaporated under reduced pressure. The residue was purified by column chromatography (SiO_2 , 75% EtOAc/hexane) to give **18** (3.99 g, 81% as a white foam): $[\alpha]_D^{-42.3}$ (*c* 1.28, CHCl_3); ${}^1\text{H}$ NMR (CDCl_3 , 500 MHz) δ 9.09 (br s, 1H, exchanged with D_2O), 8.83 (s, 1H), 8.39 (s, 1H), 8.03 (d, 2H, $J = 7.5$ Hz), 7.61 (t, 1H, $J = 7.5$ Hz), 7.53 (t, 2H, $J = 7.4$ Hz), 6.20 (d, 1H, $J = 1.9$ Hz), 4.39–4.37 (m, 2H), 4.15 (d, 1H, $J = 1.9$ Hz), 4.06 (dd, 1H, $J = 4.6, 11.3$ Hz), 3.99 (dd, 1H, $J = 3.2, 11.3$ Hz), 3.55 (s, 3H), 2.75 (br s, 1H, exchanged with D_2O), 0.81 (s, 9H), 0.11 (s, 3H), –0.03 (s, 3H); ${}^{13}\text{C}$ NMR (CDCl_3 , 125 MHz) δ 152.9, 149.7, 142.0, 134.0, 132.9, 129.0, 128.1, 127.6, 90.5, 88.3, 61.6, 58.8, 25.8, 18.2, –4.7, –5.0; MS (EI) m/z 499 (M^+). Anal. Calcd for $\text{C}_{24}\text{H}_{33}\text{N}_5\text{O}_5\text{Si}$: C, 57.69; H, 6.66; N, 14.02. Found: C, 57.73; H, 6.56; N, 14.05.

N^6 -Benzoyl-9-(3-O-*tert*-butyldimethylsilyl-2-O-methyl- β -D-xylofuranosyl)adenine (7). Trifluoroacetic acid (1.09 mL, 4.5 mmol) was added to a solution of **18** (4.51 g, 9.05 mmol), DCC (5.63 g, 27.2 mmol), and pyridine (0.91 mL, 9.1 mmol) in DMSO (50 mL) at room temperature, and the mixture was stirred for 2 h. The mixture was filtered to remove dicyclohexylurea, and the filtrate was washed with hexane (3×50 mL) and partitioned with EtOAc (200 mL) and H_2O (200 mL). The organic layer was washed with brine (20 mL), dried (Na_2SO_4), and evaporated under reduced pressure. The residue was coevaporated with toluene (3×50 mL) under reduced pressure to give crude **7** (4.77 g as a syrup, quantitative). This compound was used in the next reaction without further purification, because it was too unstable to be isolated: ${}^1\text{H}$ NMR (CDCl_3 , 500 MHz) δ 9.78 (s, 1H), 9.16 (br s, 1H, exchanged with D_2O), 8.81 (s, 1H), 8.50 (s, 1H), 8.01 (d, 2H, $J = 7.5$ Hz), 7.59 (t, 1H, $J = 7.5$ Hz), 7.51 (t, 2H, $J = 7.4$ Hz), 6.39 (s, 1H), 4.79 (d, 1H, $J = 3.7$ Hz), 4.68 (d, 1H, $J = 3.8$ Hz), 4.13 (s, 1H), 3.62 (s, 3H), 0.68 (s, 9H), 0.04 (s, 3H), –0.15 (s, 3H).

Methyl Glucuronate-D-glycal (25). A mixture of **24** (258 mg, 1.00 mmol) in MeOH (10 mL) containing NaOMe (28% in MeOH, 10 μL) was stirred for 1 h at room temperature. The mixture was neutralized

with AcOH and evaporated under reduced pressure. The resulting syrup was partitioned between EtOAc (50 mL) and H₂O (50 mL), and the organic layer was washed with H₂O (50 mL) and brine (20 mL). The organic layer was dried (Na₂SO₄) and evaporated under reduced pressure. The residue was purified by silica gel column chromatography (SiO₂, 4% MeOH/CHCl₃) to give **25** (150 mg, 86% as a white solid), which was crystallized from MeOH/CHCl₃: mp 148–151 °C; [α]_D –13.5° (c 1.00, MeOH); ¹H NMR (DMSO-*d*₆, 500 MHz) δ 6.37 (d, 1H, *J* = 6.2 Hz), 5.43 (d, 1H, *J* = 5.0 Hz, exchanged with D₂O), 4.98 (d, 1H, *J* = 4.0 Hz, exchanged with D₂O), 4.72 (t, 1H, *J* = 5.0, 6.2 Hz), 4.40 (d, 1H, *J* = 5.1 Hz), 3.82 (ddd, 1H, *J* = 5.1, 4.6, 4.0 Hz), 3.74 (ddd, 1H, *J* = 5.0, 4.6, 5.0 Hz), 3.61 (s, 3H).

Methyl (3,4-Di-*O*-tert-butylidimethylsilyl-D-glucuronate)glycol (26a). A mixture of **25** (1.74 g, 10.0 mmol), TBSCl (4.51 g, 30.0 mmol), and imidazole (4.08 g, 60.0 mmol) in DMF (50 mL) was stirred for 4 h at 100 °C. After being cooled to room temperature, MeOH (10 mL) was added. The reaction mixture was partitioned between EtOAc (200 mL) and H₂O (200 mL), and the organic layer was washed with H₂O (200 mL) and brine (50 mL), dried (Na₂SO₄), and evaporated under reduced pressure. The residue was purified by column chromatography (SiO₂, 10% EtOAc/hexane) to give **26a** (3.97 g, 99% as a colorless syrup): [α]_D –27.2° (c 1.05, CHCl₃); ¹H NMR (CDCl₃, 500 MHz) δ 6.50 (d, 1H, *J* = 6.6 Hz), 4.81 (m, 1H), 4.53 (br s, 1H), 4.23 (dd, 1H, *J* = 2.0, 4.0 Hz), 3.77 (m, 1H), 3.71 (s, 3H), 0.90 (s, 9H), 0.83 (s, 9H), 0.13 (s, 3H), 0.12 (s, 3H), 0.06 (s, 3H), 0.05 (s, 3H); ¹³C NMR (CDCl₃, 125 MHz) δ 169.7, 144.3, 101.6, 75.8, 64.5, 52.2, 25.9, 25.8, –4.4, –4.7, –4.8, –4.9; MS (FAB) *m/z* 402 (MH⁺). Anal. Calcd for C₁₉H₃₈O₅Si₂: C, 56.67; H, 9.51. Found: C, 56.55; H, 9.67.

Methyl (3,4-Di-*O*-triisopropylsilyl-D-glucuronate)glycol (26b). In a manner similar to that described for **26a**, reaction of **25** (2.00 g, 11.4 mmol) with TIPSCl (7.38 mL, 34.2 mmol) and imidazole (4.66 g, 68.4 mmol) in DMF (50 mL) and purification by column chromatography (SiO₂, 10% EtOAc/hexane) gave **26b** (5.24 g, 95% as a colorless syrup): [α]_D –24.8° (c 1.12, CHCl₃); ¹H NMR (CDCl₃, 500 MHz) δ 6.52 (d, 1H, *J* = 6.4 Hz), 4.90 (m, 1H), 4.64 (br s, 1H), 4.47 (dd, 1H, *J* = 2.0, 4.0 Hz), 3.95 (ddd, 1H, *J* = 1.2, 2.0, 4.0 Hz), 3.70 (s, 3H), 1.87–1.04 (m, 42H); ¹³C NMR (CDCl₃, 125 MHz) δ 168.8, 143.5, 101.1, 75.5, 70.7, 64.2, 52.2, 18.3, 18.2, 18.2, 18.1, 18.1, 17.9, 12.8, 12.7, 12.6, 12.6, 12.4, 12.3, 12.1; MS (FAB) *m/z* 443 (MH⁺ – *i*-Pr). Anal. Calcd for C₂₅H₅₀O₅Si₂: C, 61.68; H, 10.35. Found: C, 61.31; H, 10.11.

Methyl (3-*O*-tert-Butyldimethylsilyl-4-*O*-tert-butylidiphenylsilyl-D-glucuronate)glycol (26c). A mixture of **25** (107 mg, 0.61 mmol), TBSCl (110 mg, 0.73 mmol), and imidazole (99.7 mg, 1.46 mmol) in DMF (5 mL) was stirred for 2 h at room temperature. After MeOH (2 mL) was added, the mixture was partitioned between EtOAc (50 mL) and H₂O (50 mL), and the organic layer was washed with H₂O (20 mL) and brine (20 mL), dried (Na₂SO₄), and evaporated under reduced pressure. A mixture of the above residue, TBDPSCl (238 μ L, 0.92 mmol), and imidazole (125 mg, 1.80 mmol) in DMF (5 mL) was stirred for 4 h at 100 °C. After MeOH (2 mL) was added, the reaction mixture was partitioned between EtOAc (50 mL) and H₂O (50 mL), and the separated organic layer was washed with H₂O (20 mL) and brine (20 mL), dried (Na₂SO₄), and evaporated under reduced pressure. The residue was purified by column chromatography (SiO₂, 10% EtOAc/hexane) to give **26c** (298 mg, 93% as a colorless syrup): [α]_D –8.15° (c 1.34, CHCl₃); ¹H NMR (CDCl₃, 500 MHz) δ 7.72–7.38 (m, 10H), 6.51 (d, 1H, *J* = 6.3 Hz), 4.85 (m, 1H), 4.52 (br s, 1H), 4.32 (dd, 1H, *J* = 1.2, 5.6 Hz), 3.77 (m, 1H), 3.56 (s, 3H), 1.05 (s, 9H), 0.72 (s, 9H), –0.14 (s, 3H), –0.17 (s, 3H); ¹³C NMR (CDCl₃, 125 MHz) δ 168.6, 143.8, 136.0, 135.9, 133.7, 133.5, 130.2, 128.1, 128.0, 101.2, 74.9, 71.0, 63.6, 52.0, 27.1, 25.9, –4.5, –4.9; MS (FAB) *m/z* 469 (MH⁺ – *t*-Bu). Anal. Calcd for C₂₉H₄₂O₅Si₂: C, 66.12; H, 8.04. Found: C, 66.32; H, 7.97.

Methyl (3,4-Di-*O*-tert-butylidimethylsilyl-2,3-anhydro)-D-glucuronate (27a). A solution of **26a** (200 mg, 0.40 mmol) in CH₂Cl₂ (4 mL) was added dropwise to an acetone solution of dimethyldioxirane (ca. 0.05 M, 10 mL, 0.50 mmol), and the mixture was stirred for 2 h at room temperature. After most of the acetone was removed under reduced pressure, the residue was dissolved in CH₂Cl₂, dried (Na₂SO₄), and filtered, and the filtrate was evaporated under reduced pressure to

give **27a** (210 mg, quantitative): [α]_D –34.1° (c 1.00, CHCl₃); ¹H NMR (CDCl₃, 500 MHz) δ 5.07 (d, 1H, *J* = 2.4 Hz), 4.18 (d, 1H, *J* = 4.5 Hz), 4.09 (dd, 1H, *J* = 2.7, 8.9 Hz), 3.97 (t, 1H, *J* = 2.4 Hz), 3.74 (s, 3H), 2.92 (t, 1H, *J* = 2.4 Hz), 0.91 (s, 9H), 0.90 (s, 9H), 0.15 (s, 3H), 0.14 (s, 3H), 0.11 (s, 3H), 0.07 (s, 3H); ¹³C NMR (CDCl₃, 125 MHz) δ 170.8, 76.6, 74.5, 72.8, 69.6, 54.1, 52.5, 25.9, –1.1, –4.6, –4.7, –4.9; MS (FAB) *m/z* 419 (MH⁺). Anal. Calcd for C₁₅H₃₈O₆Si₂: C, 54.51; H, 9.15. Found: C, 54.28; H, 9.02.

Methyl (3,4-Di-*O*-triisopropylsilyl-2,3-anhydro)-D-glucuronate (27b). In a manner similar to that described for **27a**, the reaction of **26b** (1.94 g, 4.00 mmol) with an acetone solution of dimethyldioxirane (ca. 0.05 M, 100 mL, 5.00 mmol) gave **27b** (2.03 g, quantitative as a foam): ¹H NMR (CDCl₃, 500 MHz) δ 5.30 (s, 1H), 4.33 (s, 1H), 4.31 (br s, 2H), 3.72 (s, 3H), 3.09 (br s, 1H), 1.18–1.04 (m, 42H); ¹³C NMR (CDCl₃, 125 MHz) δ 169.8, 132.5, 130.8, 129.0, 128.8, 125.3, 75.5, 73.1, 67.4, 53.2, 52.0, 18.8, 18.7, 18.6, 18.3, 17.9, 14.0, 12.8, 12.7, 12.4, 12.3, 12.1; MS (FAB) *m/z* 503 (MH⁺).

Methyl (3-*O*-tert-Butyldimethylsilyl-4-*O*-tert-butylidiphenylsilyl-2,3-anhydro)-D-glucuronate (27c). In a manner similar to that described for **27a**, the reaction of **26c** (2.10 g, 4.00 mmol) with an acetone solution of dimethyldioxirane (ca. 0.05 M, 100 mL, 5.00 mmol) gave **27c** (2.21 g, quantitative): ¹H NMR (CDCl₃, 500 MHz) δ 7.74–7.36 (m, 10H), 5.12 (d, 1H, *J* = 2.4 Hz), 4.18 (br s, 2H), 4.07 (dd, 1H, *J* = 1.9, 3.8 Hz), 3.57 (s, 3H), 3.01 (d, 1H, *J* = 1.4 Hz), 1.09 (s, 9H), 0.74 (s, 9H), –0.09 (s, 3H), –0.14 (s, 3H); ¹³C NMR (CDCl₃, 125 MHz) δ 169.8, 132.5, 130.8, 129.0, 128.8, 125.3, 75.5, 73.1, 67.4, 53.2, 52.0, 18.8, 18.7, 18.6, 18.3, 17.9, 14.0, 12.8, 12.7, 12.4, 12.3, 12.1; MS (FAB) *m/z* 543 (MH⁺). Anal. Calcd for C₂₉H₄₂O₆Si₂: C, 64.17; H, 7.80. Found: C, 64.68; H, 7.76.

Methyl 3,4-Di-*O*-tert-butylidimethylsilyl-1-phenylthio-D-glucuronate (28a). Boron trifluoride diethyl ether complex (49 μ L, 0.4 mmol) was added dropwise to a mixture of **27a** (1.67 g, 4.00 mmol) and PhSH (4.11 mL, 40.0 mmol) in CH₂Cl₂ (40 mL), and the mixture was stirred for 30 min at –40 °C. The mixture was washed with saturated aqueous NaHCO₃ (40 mL), H₂O (50 mL), and brine (50 mL), dried (Na₂SO₄), and evaporated under reduced pressure. The residue was purified by column chromatography (SiO₂, 75% benzene/hexane) to give **28a** as an inseparable anomeric mixture (1.44 g, 80% as a colorless syrup, the ratio of α/β = 39/61): MS (FAB) *m/z* 528 (M⁺). Anal. Calcd for C₂₅H₄₄O₆SSi₂: C, 56.78; H, 8.39. Found: C, 56.93; H, 8.09. For β -anomer: ¹H NMR (CDCl₃, 500 MHz) δ 7.57 (d, 2H, *J* = 7.5 Hz), 7.33–7.18 (t, 3H), 5.16 (d, 1H, *J* = 5.8 Hz), 4.30 (t, 1H, *J* = 3.8 Hz), 4.27 (d, 1H, *J* = 3.1 Hz), 3.85 (t, 1H, *J* = 4.3 Hz), 3.67 (s, 3H), 3.22 (d, 1H, *J* = 7.7 Hz, exchanged with D₂O), 0.89 (s, 9H), 0.88 (s, 9H), 0.18 (s, 3H), 0.17 (s, 3H), 0.15 (s, 3H), 0.11 (s, 3H); ¹³C NMR (CDCl₃, 125 MHz) δ 169.9, 135.5, 133.1, 129.4, 129.0, 87.1, 72.8, 71.8, 70.7, 64.4, 52.1, 26.2, 25.9, 18.6, 18.1, –4.1, –4.2, –4.8, –4.9. For α -anomer: ¹H NMR (CDCl₃, 500 MHz) δ 7.71 (d, 2H, *J* = 7.7 Hz), 7.33–7.18 (t, 3H), 5.94 (s, 1H), 4.48 (s, 1H), 4.20 (t, 1H, *J* = 1.4 Hz), 3.98 (t, 1H, *J* = 3.4 Hz), 3.95 (s, 1H), 3.73 (m, 2H), 3.72 (s, 3H), 0.94 (s, 9H), 0.85 (s, 9H), 0.18 (s, 3H), 0.17 (s, 3H), 0.07 (s, 3H), 0.06 (s, 3H); ¹³C NMR (CDCl₃, 125 MHz) δ 169.7, 143.7, 133.1, 127.3, 126.4, 81.7, 77.8, 75.6, 73.1, 72.2, 69.9, 26.0, 25.8, 18.2, 11.7, –4.3, –4.5, –4.5, –4.6.

Methyl 3,4-Di-*O*-triisopropylsilyl-1-phenylthio-D-glucuronate (28b). In a manner similar to that described for **28a**, reaction of **27b** (2.03 g, 4.00 mmol) with BF₃·OEt₂ (49 μ L, 0.4 mmol) and PhSH (4.11 mL, 40.0 mmol) in CH₂Cl₂ (40 mL) and purification by column chromatography (SiO₂, 75% benzene/hexane) gave **28b** as an inseparable anomeric mixture (1.53 g, 63% as a colorless syrup, the ratio of α/β = 60/40): MS (FAB) *m/z* 613 (M⁺). Anal. Calcd for C₃₁H₅₆O₆SSi₂: C, 60.74; H, 9.21. Found: C, 60.42; H, 9.53. For α -anomer: ¹H NMR (CDCl₃, 500 MHz) δ 7.59 (d, 2H, *J* = 7.3 Hz), 7.32–7.19 (m, 3H), 5.35 (d, 1H, *J* = 5.8 Hz), 4.68 (d, 1H, *J* = 2.3 Hz), 4.54 (s, 1H), 4.12 (d, 1H, *J* = 6.5 Hz), 3.82 (m, 1H), 3.66 (s, 3H), 3.20 (d, 1H, *J* = 8.8 Hz, exchanged with D₂O), 1.12–1.04 (m, 42H); ¹³C NMR (CDCl₃, 125 MHz) δ 171.1, 136.6, 132.4, 130.0, 127.4, 87.0, 74.6, 73.6, 72.3, 71.2, 53.1, 19.5, 19.3, 19.2, 19.1, 13.6, 13.4. For β -anomer: ¹H NMR (CDCl₃, 500 MHz) δ 7.70 (d, 2H, *J* = 7.0 Hz), 7.31–7.22 (m, 3H), 5.99 (s, 1H), 4.58 (s, 1H), 4.43 (m, 1H), 4.20 (t, 1H, *J* = 3.3 Hz), 4.05 (d, 1H, *J* = 10.1 Hz, exchanged with D₂O), 3.82 (m, 1H), 3.69 (s,

3H), 1.12–1.00 (m, 42H); ¹³C NMR (CDCl₃, 125 MHz) δ 170.7, 144.6, 136.9, 130.6, 130.0, 128.2, 102.2, 82.9, 76.6, 73.5, 71.8, 65.2, 19.3, 19.2, 19.1, 18.9, 13.7, 13.5.

Methyl 3-*O*-tert-Butyldimethylsilyl-4-*O*-tert-butylidiphenylsilyl-1-phenylthio-*D*-glucuronate (28c). In a manner similar to that described for **28a**, reaction of **27c** (2.71 g, 4.00 mmol) with BF₃·OEt₂ (49 μL, 0.4 mmol) and PhSH (4.11 mL, 40.0 mmol) in CH₂Cl₂ (40 mL) and purification by column chromatography (SiO₂, 75% benzene/hexane) gave **28c** as an inseparable anomeric mixture (1.96 g, 75% as a colorless syrup, the ratio of α/β = 72/28): MS (FAB) *m/z* 652 (M⁺). Anal. Calcd for C₃₅H₄₈O₆SSi₂·0.5benzene: C, 65.95; H, 7.42. Found: C, 65.98; H, 7.41. For α-anomer: ¹H NMR (CDCl₃, 500 MHz) δ 7.89–7.46 (m, 15H), 5.67 (d, 1H, *J* = 5.0 Hz), 4.64 (d, 1H, *J* = 1.7 Hz), 4.55 (s, 1H), 4.17 (t, 1H, *J* = 3.0 Hz), 3.99 (m, 1H), 3.72 (d, 1H, *J* = 7.8 Hz, exchanged with D₂O), 3.64 (s, 3H), 1.23 (s, 9H), 0.94 (s, 9H), 0.40 (s, 3H), 0.00 (s, 3H); ¹³C NMR (CDCl₃, 125 MHz) δ 169.7, 1362–126.3, 86.5, 73.1, 72.4, 71.4, 51.8, 27.2, 27.2, 25.9, 19.3, 19.2, 18.3, –5.0, –5.1. For β-anomer: ¹H NMR (CDCl₃, 500 MHz) δ 7.89–7.46 (m, 15H), 6.10 (s, 1H), 4.57 (s, 1H), 4.37 (t, 1H, *J* = 1.6 Hz), 4.24 (d, 1H, *J* = 11.9 Hz, exchanged with D₂O), 4.20 (t, 1H, *J* = 3.4 Hz), 3.99 (m, 1H), 3.68 (s, 3H), 1.30 (s, 9H), 0.89 (s, 9H), 0.05 (s, 3H), –0.03 (s, 3H); ¹³C NMR (CDCl₃, 125 MHz) δ 169.4, 1362–126.3, 81.6, 76.1, 72.2, 71.2, 70.0, 26.8, 25.8, 19.3, 18.1, –5.0, –5.2.

Methyl 3,4-Di-*O*-tert-butylidimethylsilyl-1-phenylthio-2-ulos-*D*-glucuronate (29a). A mixture of **28a** (1.31 g, 2.48 mmol, the ratio of α/β = 39/61) and Dess–Martin periodinane (1.26 g, 2.98 mmol) in CH₂Cl₂ (25 mL) was stirred for 3 h at room temperature. After a mixture of saturated aqueous NaHCO₃ and saturated aqueous Na₂S₂O₃ (5:1) was added, the mixture was vigorously stirred until the organic layer turned clear. The separated organic layer was washed with H₂O (50 mL) and brine (50 mL), dried (Na₂SO₄), and evaporated under reduced pressure. The residue was purified by column chromatography (SiO₂, 50% benzene/hexane) to give **29a** as an inseparable anomeric mixture (1.23 g, 94% as a colorless syrup, the ratio of α/β = 39/61): MS (FAB) *m/z* 527 (MH⁺). Anal. Calcd for C₂₅H₄₂O₆SSi₂: C, 57.00; H, 8.04. Found: C, 56.81; H, 7.80. For β-anomer: ¹H NMR (CDCl₃, 500 MHz) δ 7.50 (d, 2H, *J* = 7.0 Hz), 7.23–7.15 (m, 3H), 5.47 (s, 1H), 4.45 (dd, 1H, *J* = 3.3, 5.4 Hz), 4.33 (d, 1H, *J* = 3.3 Hz), 3.96 (d, 1H, *J* = 5.4 Hz), 3.69 (s, 3H), 0.79 (s, 9H), 0.76 (s, 9H), 0.01 (s, 3H), 0.00 (s, 3H), –0.01 (s, 3H), –0.08 (s, 3H); ¹³C NMR (CDCl₃, 125 MHz) δ 199.2, 169.1, 133.4, 132.2, 128.9, 128.1, 127.7, 86.9, 78.9, 76.3, 74.0, 52.5, 26.0, 25.8, –4.8, –4.9, –5.1. For α-anomer: ¹H NMR (CDCl₃, 500 MHz) δ 7.45 (d, 2H, *J* = 7.0 Hz), 7.23–7.15 (m, 3H), 5.75 (s, 1H), 4.68 (d, 1H, *J* = 5.8 Hz), 4.17 (dd, 1H, *J* = 5.8, 12.8 Hz), 4.17 (d, 1H, *J* = 12.8 Hz), 3.70 (s, 3H), 0.84 (s, 9H), 0.80 (s, 9H), 0.03 (s, 3H), –0.04 (s, 3H), –0.05 (s, 3H), –0.09 (s, 3H); ¹³C NMR (CDCl₃, 125 MHz) δ 198.1, 169.1, 132.8, 132.1, 129.2, 128.3, 88.2, 78.1, 75.5, 75.4, 52.4, 25.8, 25.7, –3.8, –4.1, –4.5.

Methyl 3,4-Di-*O*-triisopropylsilyl-1-phenylthio-2-ulos-*D*-glucuronate (29b). In a manner similar to that described for **29a**, the reaction of **28b** (1.39 g, 2.28 mmol, the ratio of α/β = 60/40) with Dess–Martin periodinane (1.16 g, 2.74 mmol) in CH₂Cl₂ (25 mL) and purification by column chromatography (SiO₂, 50% benzene/hexane) gave **29b** as an inseparable anomeric mixture (1.12 g, 81% as a colorless syrup, the ratio of α/β = 60/40): MS (FAB) *m/z* 611 (M⁺). Anal. Calcd for C₃₁H₅₄O₆SSi₂: C, 60.94; H, 8.91. Found: C, 60.83; H, 8.83. For α-anomer: ¹H NMR (CDCl₃, 500 MHz) δ 7.62 (d, 2H, *J* = 7.0 Hz), 7.31–7.22 (m, 3H), 5.65 (s, 1H), 4.92 (d, 1H, *J* = 4.3 Hz), 4.6 (s, 1H), 4.15 (d, 1H, *J* = 4.3 Hz), 3.79 (s, 3H), 1.12–1.00 (m, 42H); ¹³C NMR (CDCl₃, 125 MHz) δ 199.2, 169.1, 133.4, 132.2, 129.2, 128.3, 127.7, 86.9, 78.9, 76.3, 74.0, 52.5, 25.9, 25.8. For β-anomer: ¹H NMR (CDCl₃, 500 MHz) δ 7.64 (d, 2H, *J* = 7.0 Hz), 7.31–7.22 (m, 3H), 6.42 (s, 1H), 4.85 (d, 1H, *J* = 4.1 Hz), 4.70 (s, 1H), 4.19 (d, 1H, *J* = 4.1 Hz), 3.78 (s, 3H), 1.12–1.00 (m, 42H); ¹³C NMR (CDCl₃, 125 MHz) δ 198.0, 169.0, 132.8, 132.1129.2, 128.3, 128.1, 88.2, 78.1, 75.5, 75.4, 52.4, 25.8, 25.7.

Methyl 3-*O*-tert-Butyldimethylsilyl-4-*O*-tert-butylidiphenylsilyl-1-phenylthio-2-ulos-*D*-glucuronate (29c). In a manner similar to that described for **28a**, the reaction of **28c** (1.78 g, 2.74 mmol, the ratio of α/β = 72/28) with Dess–Martin periodinane (1.39 g, 3.29 mmol) in CH₂Cl₂ (30 mL) and purification by column chromatography (SiO₂,

50% benzene/hexane) gave an inseparable anomeric mixture of **29c** (1.75 g, 99% as a colorless syrup, the ratio of α/β = 74/26): MS (FAB) *m/z* 651 (MH⁺). Anal. Calcd for C₃₅H₄₆O₆SSi₂·1/6benzene: C, 65.12; H, 7.13. Found: C, 65.22; H, 7.26. For α-anomer: ¹H NMR (CDCl₃, 500 MHz) δ 7.73–7.26 (m, 15H), 5.74 (s, 1H), 4.70 (d, 1H, *J* = 4.0 Hz), 4.56 (s, 1H), 3.87 (d, 1H, *J* = 4.0 Hz), 3.65 (s, 3H), 0.95 (s, 9H), 0.89 (s, 9H), –0.18 (s, 3H), –0.19 (s, 3H); ¹³C NMR (CDCl₃, 125 MHz) δ 199.0, 168.6, 136.1, 135.9, 135.7, 132.2, 130.1, 129.0, 127.8, 127.7, 86.4, 77.9, 74.1, 73.3, 52.1, 26.8, 26.7. For β-anomer: ¹H NMR (CDCl₃, 500 MHz) δ 7.73–7.26 (m, 15H), 6.41 (s, 1H), 4.66 (s, 1H), 4.58 (d, 1H, *J* = 4.0 Hz), 3.89 (d, 1H, *J* = 4.0 Hz), 3.62 (s, 3H), 0.93 (s, 9H), 0.91 (s, 9H), –0.14 (s, 3H), –0.21 (s, 3H); ¹³C NMR (CDCl₃, 125 MHz) δ 193.4, 164.7, 135.8, 135.7, 135.6, 133.9, 132.2, 131.9, 129.8, 127.6, 127.5, 86.3, 76.2, 76.6, 76.1, 50.9, 26.7, 26.5.

N⁶-Benzoyl-9-[methyl 6,10-Anhydro-3,8,9-tri-*O*-(tert-butylidimethylsilyl)-2-*O*-methyl-α-*D*-arabino-1-ido-7-undeculofuranuronyl]adenine (30a-α) and N⁶-Benzoyl-9-[methyl 6,10-Anhydro-3,8,9-tri-*O*-(tert-butylidimethylsilyl)-2-*O*-methyl-β-*D*-arabino-*D*-gluco-7-undeculofuranuronate]adenine (30a-β). A THF solution of SmI₂ (0.1 M, 7.0 mL, 0.70 mmol) was cooled to –40 °C, and then **29a** (168 mg, 0.32 mmol) in THF (3 mL) was added dropwise to the mixture. After the TLC analysis indicated the disappearance of **29a**, oxygen gas was passed through the mixture. Then, **7** (160 mg, 0.32 mmol) in THF (3 mL) was added dropwise to the above mixture, and the whole was stirred for 15 min at the same temperature. After the mixture was allowed to warm to room temperature, saturated aqueous NH₄Cl was added. The mixture was filtrated through a Celite pad, the filtrate was partitioned between EtOAc (50 mL) and H₂O (50 mL), and the separated organic layer was washed with saturated aqueous NaHCO₃ (50 mL) and brine (30 mL), dried (Na₂SO₄), and evaporated under reduced pressure. The residue was purified by flash column chromatography (SiO₂, 40% EtOAc/hexane) to give **30a-α** (22.8 mg, 16% as a white foam) and **30a-β** (123 mg, 42% as a white foam). For α-anomer: [α]_D –45.0° (c 1.10, CHCl₃); ¹H NMR (CDCl₃, 500 MHz) δ 9.02 (br s, 1H, exchanged with D₂O), 8.83 (s, 1H), 8.40 (s, 1H), 8.03 (d, 2H, *J* = 7.6 Hz), 7.61 (t, 1H, *J* = 7.4 Hz), 7.53 (t, 2H, *J* = 7.6 Hz), 6.23 (s, 1H), 4.56 (m, 2H), 4.52 (m, 3H), 3.99 (br s, 1H), 3.86 (d, 1H, *J* = 2.8 Hz), 3.80 (s, 3H), 3.71 (d, 1H, *J* = 5.1 Hz), 3.61 (s, 3H), 0.89 (s, 9H), 0.85 (s, 9H), 0.74 (s, 9H), 0.13 (s, 3H), 0.11 (s, 3H), 0.10 (s, 3H), 0.07 (s, 3H), 0.02 (s, 3H), –0.08 (s, 3H); ¹³C NMR (CDCl₃, 125 MHz) δ 203.0, 170.1, 152.4, 150.7, 149.1, 141.9, 133.8, 132.6, 132.5, 128.7, 127.8, 122.8, 90.6, 88.5, 82.1, 80.7, 77.9, 75.7, 75.1, 73.4, 69.9, 58.2, 52.4, 25.7, 25.6, 25.5, 18.3, 18.0, –4.7, –4.8, –4.9, –5.1; MS (FAB) *m/z* 917 (MH⁺); exact MS (FAB) calcd for C₄₃H₇₀N₅O₁₁Si₃ 916.4379, found 916.4398. Anal. Calcd for C₄₃H₆₉N₅O₁₁Si₃: C, 56.36; H, 7.59; N, 7.64. Found: C, 55.90; H, 7.47; N, 7.69. For β-anomer: [α]_D +5.08° (c 1.15, CHCl₃); ¹H NMR (CDCl₃, 500 MHz) δ 9.00 (br s, 1H, exchanged with D₂O), 8.82 (s, 1H), 8.65 (s, 1H), 7.92 (d, 2H, *J* = 7.5 Hz), 7.61 (t, 1H, *J* = 7.2 Hz), 7.53 (t, 2H, *J* = 7.3 Hz), 6.23 (s, 1H, *J* = 3.6 Hz), 4.97 (d, 1H, *J* = 7.1 Hz), 4.47 (d, 1H, *J* = 7.1 Hz), 4.56 (dd, 2H, *J* = 3.2, 6.6 Hz), 4.37 (m, 2H), 4.26 (d, 1H, *J* = 4.5 Hz), 3.72 (s, 3H), 3.64 (d, 1H, *J* = 4.4 Hz), 3.44 (s, 3H), 0.89 (s, 9H), 0.85 (s, 9H), 0.84 (s, 9H), 0.13 (s, 3H), 0.12 (s, 3H), 0.11 (s, 3H), 0.07 (s, 3H), 0.04 (s, 3H), 0.02 (s, 3H); ¹³C NMR (CDCl₃, 125 MHz) δ 205.6, 169.0, 152.2, 151.0, 149.1, 142.0, 133.5, 132.3, 128.5, 127.7, 122.7, 89.2, 86.7, 80.3, 77.3, 77.2, 77.0, 76.7, 76.5, 76.1, 75.8, 75.3, 74.2, 68.0, 58.4, 52.0, 25.6, 25.5, 25.4, 18.1, 17.8, 17.7, –4.8, –4.8, –5.0, –5.0, –5.1, –5.2; MS (FAB) *m/z* 917 (MH⁺); exact MS (FAB) calcd for C₄₃H₇₀N₅O₁₁Si₃ 916.4379, found 916.4352. Anal. Calcd for C₄₃H₆₉N₅O₁₁Si₃: C, 56.36; H, 7.59; N, 7.64. Found: C, 56.36; H, 7.50; N, 7.61.

N⁶-Benzoyl-9-[methyl 6,10-Anhydro-3-*O*-(tert-butylidimethylsilyl)-2-*O*-methyl-8,9-di-*O*-(triisopropylsilyl)-α-*D*-arabino-1-ido-7-undeculofuranuronyl]adenine (30b-α) and N⁶-Benzoyl-9-[methyl 6,10-Anhydro-3-*O*-(tert-butylidimethylsilyl)-2-*O*-methyl-8,9-di-*O*-(triisopropylsilyl)-β-*D*-arabino-*D*-gluco-7-undeculofuranuronate]adenine (30b-β). In a manner similar to that described for **30a**, from **29b** (196 mg, 0.32 mmol), **30b-α** (122 mg, 38% as a white foam) and **30b-β** (103 mg, 33% as a white foam) were obtained after purification by flash column chromatography (SiO₂, 40% EtOAc/hexane). For α-anomer: [α]_D –32.7° (c 1.06, CHCl₃); ¹H NMR (CDCl₃, 500 MHz) δ

9.08 (br s, 1H, exchanged with D₂O), 8.83 (s, 1H), 8.39 (s, 1H), 8.03 (d, 2H, *J* = 7.5 Hz), 7.61 (t, 1H, *J* = 7.4 Hz), 7.53 (t, 2H, *J* = 7.8 Hz), 6.22 (s, 1H), 4.84 (d, 1H, *J* = 4.3 Hz), 4.61 (s, 1H), 4.58 (br s, 3H), 4.54 (s, 1H), 4.03 (d, 1H, *J* = 4.3 z), 3.93 (br s, 1H), 3.79 (s, 3H), 3.61 (s, 3H), 3.57 (br s, 1H, exchanged with D₂O), 1.05–1.01 (m, 42H), 0.73 (s, 9H), 0.13 (s, 3H), –0.96 (s, 3H); ¹³C NMR (CDCl₃, 125 MHz) δ 203.4, 169.9, 164.5, 152.5, 149.0, 141.9, 133.8, 132.5, 128.7, 127.7, 90.5, 88.6, 82.1, 80.3, 77.9, 77.2, 75.1, 74.1, 69.8, 58.2, 52.3, 25.5, 17.9, 17.7, 12.1, –5.1, –5.2; MS (FAB) *m/z* 1001 (MH⁺); exact MS (FAB) calcd for C₄₉H₈₁N₅O₁₁Si₃ 1000.5318, found 1000.5320. Anal. Calcd for C₄₉H₈₁N₅O₁₁Si₃: C, 58.83; H, 8.16; N, 7.00. Found: C, 58.62; H, 8.09; N, 6.75. For β-anomer: [α]_D 1 + 12.1° (*c* 0.94, CHCl₃); ¹H NMR (CDCl₃, 500 MHz) δ 9.01 (br s, 1H, exchanged with D₂O), 8.82 (s, 1H), 8.72 (s, 1H), 8.02 (d, 2H, *J* = 7.1 Hz), 7.61 (t, 1H, *J* = 7.4 Hz), 7.53 (t, 2H, *J* = 7.4 Hz), 6.24 (s, 1H, *J* = 4.9 Hz), 5.16 (d, 1H, *J* = 7.9 Hz), 4.82 (dd, 1H, *J* = 1.7, 4.0 Hz), 4.62 (dd, 1H, *J* = 1.4, 6.8 Hz), 4.54 (d, 1H, *J* = 6.2 Hz), 4.39 (br s, 1H), 4.32 (m, 2H), 4.10 (d, 1H, *J* = 4.0 z), 3.70 (s, 3H), 3.37 (s, 3H), 3.29 (br s, 1H, exchanged with D₂O), 1.06–1.00 (m, 42H), 0.87 (s, 9H), 0.11 (s, 3H), 0.06 (s, 3H); ¹³C NMR (CDCl₃, 125 MHz) δ 206.0, 168.9, 164.4, 152.5, 151.3, 149.1, 142.3, 133.8, 132.5, 128.7, 127.7, 122.7, 88.8, 86.4, 78.9, 76.5, 76.2, 75.6, 75.5, 72.1, 67.6, 58.7, 52.0, 25.6, 17.9, 17.8, 17.7, 17.6, 12.2, 12.0, –4.8, –5.0; MS (FAB) *m/z* 1001 (MH⁺); exact MS (FAB) calcd for C₄₉H₈₂N₅O₁₁Si₃ 1000.5318, found 1000.5390. Anal. Calcd for C₄₉H₈₁N₅O₁₁Si₃: C, 58.83; H, 8.16; N, 7.00. Found: C, 58.74; H, 8.02; N, 6.99.

N⁶-Benzoyl-9-[methyl 6,10-Anhydro-3,8-*O*-(*tert*-butyldimethylsilyl)-2-*O*-methyl-9-*O*-(*tert*-butyldiphenylsilyl)-α-*D*-arabino-*L*-ido-7-undeculofuranuronyl]adenine (30c-α) and N⁶-Benzoyl-9-[methyl 6,10-Anhydro-3,8-*O*-(*tert*-butyldimethylsilyl)-2-*O*-methyl-9-*O*-(*tert*-butyldiphenylsilyl)-β-*D*-arabino-*D*-gluco-7-undeculofuranuronate]adenine (30c-β). In a manner similar to that described for 30a, from 29c (300 mg, 0.46 mmol), 30c-α (198 mg, 41% as a white foam) and 30c-β (135 mg, 29% as a white foam) were obtained after purification by flash column chromatography (SiO₂, 40% EtOAc/hexane). For α-anomer: [α]_D –36.3° (*c* 1.15, CHCl₃); ¹H NMR (CDCl₃, 500 MHz) δ 9.06 (br s, 1H, exchanged with D₂O), 8.84 (s, 1H), 8.43 (s, 1H), 8.03 (d, 2H, *J* = 7.3 Hz), 7.65–7.39 (m, 13H), 6.26 (s, 1H), 4.74 (d, 1H, *J* = 2.1 Hz), 4.57 (m, 5H), 3.98 (br s, 2H), 3.68 (d, 1H, *J* = 4.2 z), 3.64 (s, 3H), 3.60 (s, 3H), 1.02 (s, 9H), 0.74 (s, 9H), 0.68 (s, 9H), 0.14 (s, 3H), –0.08 (s, 3H), –0.17 (s, 3H), –0.23 (s, 3H); ¹³C NMR (CDCl₃, 125 MHz) δ 203.3, 170.2, 152.5, 150.7, 149.1, 141.9, 135.7, 133.8, 132.5, 132.1, 132.0, 130.3, 130.1, 128.7, 127.9, 127.8, 127.7, 122.8, 90.7, 88.5, 82.1, 80.8, 77.2, 75.0, 73.3, 73.1, 70.2, 58.2, 52.2, 26.8, 25.6, 25.4, 19.1, 18.1, 18.0, –5.1, –5.2, –5.6; MS (FAB) *m/z* 1041 (MH⁺); exact MS (FAB) calcd for C₅₃H₇₄N₅O₁₁Si₃ 1040.4692, found 1040.4700. Anal. Calcd for C₅₃H₇₃N₅O₁₁Si₃·EtOAc: C, 60.66; H, 7.23; N, 6.21. Found: C, 60.35; H, 7.05; N, 6.25. For β-anomer: [α]_D +11.4° (*c* 1.06, CHCl₃); ¹H NMR (CDCl₃, 500 MHz) δ 9.03 (br s, 1H, exchanged with D₂O), 8.82 (s, 1H), 8.72 (s, 1H), 8.02 (d, 2H, *J* = 7.3 Hz), 7.60–7.38 (m, 13H), 6.25 (d, 1H, *J* = 4.4 Hz), 5.06 (d, 1H, *J* = 7.9 Hz), 4.69 (d, 1H, *J* = 6.5 Hz), 4.55 (m, 3H), 4.41 (d, 1H, *J* = 6.7 Hz), 4.32 (br s, 2H), 3.84 (d, 1H, *J* = 4.0 Hz), 3.57 (s, 3H), 3.39 (s, 3H), 1.06 (s, 9H), 0.88 (s, 9H), 0.69 (s, 9H), 0.15 (s, 3H), 0.07 (s, 3H), –0.16 (s, 3H), –0.17 (s, 3H); ¹³C NMR (CDCl₃, 125 MHz) δ 206.1, 168.8, 152.5, 151.4, 149.2, 142.3, 135.6, 135.5, 133.8, 132.5, 132.2, 130.2, 130.1, 128.7, 127.9, 127.8, 127.7, 122.6, 89.0, 86.6, 79.3, 76.7, 75.7, 75.5, 75.0, 67.8, 58.7, 51.9, 89.0, 86.6, 79.3, 77.3, 77.0, 76.7, 75.7, 75.5, 75.0, 72.5, 67.8, 58.7, 51.9, 26.8, 25.7, 25.4, 19.2, 18.0, 17.9, –4.7, –4.8, –5.2, –5.3; MS (FAB) *m/z* 1041 (MH⁺); exact MS (FAB) calcd for C₅₃H₇₄N₅O₁₁Si₃ 1040.4692, found 1040.4680. Anal. Calcd for C₅₃H₇₃N₅O₁₁Si₃·EtOAc: C, 60.66; H, 7.23; N, 6.21. Found: C, 60.47; H, 6.97; N, 6.50.

N⁶-Benzoyl-9-[methyl 6,10-Anhydro-3,8,9-tri-*O*-(*tert*-butyldimethylsilyl)-6-dehydro-5-deoxy-2-*O*-methyl-α-*D*-arabino-*L*-ido-7-undeculofuranuronyl]adenine (31a). A mixture of 30a (60 mg, 66 μmol) and Burgess's inner salt (96 mg, 396 μmol) in toluene (5 mL) was heated under reflux for 1 h. After being cooled to room temperature, the mixture was evaporated under reduced pressure. The residue in EtOAc (30 mL) was washed with H₂O (3 × 20 mL) and brine (10 mL). The organic layer was dried (Na₂SO₄) and evaporated under

reduced pressure. The residue was purified by flash column chromatography (SiO₂, 40% EtOAc/hexane) to give 31a (28 mg, 66% as a pale yellow foam): [α]_D –7.55° (*c* 1.02, CHCl₃); ¹H NMR (CDCl₃, 500 MHz) δ 9.04 (br s, 1H, exchanged with D₂O), 8.84 (s, 1H), 8.41 (s, 1H), 8.03 (d, 2H, *J* = 7.5 Hz), 7.61 (t, 1H, *J* = 7.5 Hz), 7.54 (t, 2H, *J* = 7.5 Hz), 6.32 (s, 1H), 6.08 (d, 1H, *J* = 8.1 Hz), 5.36 (dd, 1H, *J* = 2.9, 8.1 Hz), 4.65 (d, 1H, *J* = 1.7 Hz), 4.56 (dd, 1H, *J* = 1.7, 3.8 Hz), 4.28 (d, 1H, *J* = 2.8 Hz), 4.12 (d, 1H, *J* = 1.7 Hz), 3.99 (d, 1H, *J* = 3.9 Hz), 3.77 (s, 3H), 3.61 (s, 3H), 0.89 (s, 9H), 0.86 (s, 9H), 0.76 (s, 9H), 0.15 (s, 3H), 0.13 (s, 3H), 0.10 (s, 3H), 0.07 (s, 3H), 0.02 (s, 3H), –0.09 (s, 3H); ¹³C NMR (CDCl₃, 125 MHz) δ 189.2, 168.5, 152.5, 150.8, 149.2, 147.5, 141.9, 133.8, 132.6, 128.7, 127.8, 122.6, 111.3, 91.4, 88.1, 79.6, 76.5, 76.4, 72.8, 72.5, 58.4, 52.4, 25.6, 25.6, 25.5, 18.4, 18.0, –4.8, –4.9, –5.0, –5.0, –5.1, –5.2; MS (FAB) *m/z* 899 (MH⁺); exact MS (FAB) calcd for C₄₃H₆₈N₅O₁₀Si₃ 898.4273, found 898.4302. Anal. Calcd for C₄₃H₆₇N₅O₁₀Si₃·H₂O: C, 56.36; H, 7.59; N, 7.64. Found: C, 56.16; H, 7.39; N, 7.68.

N⁶-Benzoyl-9-[methyl 6,10-Anhydro-3-*O*-(*tert*-butyldimethylsilyl)-6-dehydro-5-deoxy-8,9-di-*O*-(triisopropylsilyl)-2-*O*-methyl-α-*D*-arabino-*L*-ido-7-undeculofuranuronyl]adenine (31b). In a manner similar to that described for 31a, the reaction of 30b (60 mg, 60 μmol) with Burgess's inner salt (87 mg, 360 μmol) in toluene (5 mL) and purification by flash silica gel column chromatography (SiO₂, 40% EtOAc/hexane) gave 31b (41 mg, 70% as a pale yellow foam): [α]_D –13.6° (*c* 1.02, CHCl₃); ¹H NMR (CDCl₃, 500 MHz) δ 9.04 (br s, 1H, exchanged with D₂O), 8.84 (s, 1H), 8.40 (s, 1H), 8.03 (d, 2H, *J* = 7.6 Hz), 7.61 (t, 1H, *J* = 7.5 Hz), 7.53 (t, 2H, *J* = 7.6 Hz), 6.31 (s, 1H), 6.04 (d, 1H, *J* = 8.2 Hz), 5.39 (dd, 1H, *J* = 2.9, 8.2 Hz), 4.83 (dd, 1H, *J* = 1.7, 3.6 Hz), 4.75 (s, 1H), 4.26 (d, 1H, *J* = 2.9 Hz), 4.13 (d, 1H, *J* = 3.9 Hz), 3.99 (s, 1H), 3.76 (s, 3H), 3.63 (s, 3H), 1.14–1.02 (m, 42H), 0.76 (s, 9H), 0.04 (s, 3H), –0.10 (s, 3H); ¹³C NMR (CDCl₃, 125 MHz) δ 189.2, 168.5, 152.5, 150.8, 149.2, 147.5, 141.9, 133.8, 132.6, 128.7, 127.8, 122.6, 111.3, 91.4, 88.1, 79.6, 76.5, 76.4, 72.8, 72.5, 58.4, 52.4, 25.6, 25.6, 25.5, 18.4, 18.0, –4.8, –4.9, –5.0, –5.0, –5.1, –5.2; MS (FAB) *m/z* 983 (MH⁺); exact MS (FAB) calcd for C₄₉H₈₀N₅O₁₀Si₃ 982.5212, found 982.5192. Anal. Calcd for C₄₉H₇₉N₅O₁₀Si₃·0.5EtOAc: C, 59.67; H, 8.15; N, 6.82. Found: C, 59.26; H, 7.76; N, 7.16.

N⁶-Benzoyl-9-[methyl 6,10-Anhydro-3,8-di-*O*-(*tert*-butyldimethylsilyl)-6-dehydro-5-deoxy-9-*O*-(*tert*-butyldiphenylsilyl)-2-*O*-methyl-α-*D*-arabino-*L*-ido-7-undeculofuranuronyl]adenine (31c). In a manner similar to that described for 31a, the reaction of 30c (60 mg, 66 μmol) with Burgess's inner salt (96 mg, 396 μmol) in toluene (5 mL) and purification by flash silica gel column chromatography (SiO₂, 40% EtOAc/hexane) gave 31c (28 mg, 79% as a pale yellow foam): [α]_D –22.0° (*c* 0.68, CHCl₃); ¹H NMR (CDCl₃, 500 MHz) δ 9.12 (br s, 1H, exchanged with D₂O), 8.84 (s, 1H), 8.43 (s, 1H), 8.03 (d, 2H, *J* = 7.5 Hz), 7.61 (t, 1H, *J* = 7.5 Hz), 7.54 (t, 2H, *J* = 7.5 Hz), 7.49–7.40 (m, 10H), 6.33 (s, 1H), 6.11 (d, 1H, *J* = 8.3 Hz), 5.42 (dd, 1H, *J* = 2.9, 8.3 Hz), 4.68 (d, 1H, *J* = 1.4 Hz), 4.55 (dd, 1H, *J* = 2.0, 3.6 Hz), 4.32 (d, 1H, *J* = 2.9 Hz), 4.01 (s, 1H), 3.83 (d, 1H, *J* = 3.6 Hz), 3.65 (s, 3H), 3.64 (s, 3H), 1.06 (s, 9H), 0.78 (s, 9H), 0.69 (s, 9H), 0.09 (s, 3H), 0.07 (s, 3H), –0.14 (s, 3H), –0.15 (s, 3H); ¹³C NMR (CDCl₃, 125 MHz) δ 181.3, 169.6, 169.3, 163.0, 139.7, 134.6, 132.0, 129.3, 128.1, 111.4, 102.4, 101.7, 100.1, 88.6, 87.2, 84.9, 76.4, 76.2, 75.9, 71.9, 69.5, 68.4, 63.8, 36.0, 30.3, 27.9, 26.0, 21.4, 20.9, –4.0, –4.2, –4.5, –4.8; MS (FAB) *m/z* 1023 (MH⁺); exact MS (FAB) calcd for C₅₃H₇₂N₅O₁₀Si₃ 1022.4586, found 1022.4579. Anal. Calcd for C₅₃H₇₁N₅O₁₀Si₃: C, 62.26; H, 7.00; N, 6.85. Found: C, 62.54; H, 6.77; N, 6.32.

Herbicidin B (1b). A mixture of 31c (50 mg, 55 μmol), HCO₂NH₄ (173 mg, 5.5 mmol), and Pd-on-carbon (20%, 15 mg) in MeOH (5 mL) was stirred for 24 h at room temperature. The mixture was filtrated through a Celite pad, and the filtrate was evaporated under reduced pressure. The residue was partitioned between EtOAc (5 mL) and H₂O (2 mL), and the separated organic layer was washed with H₂O (3 mL) and brine (3 mL). The organic layer was dried (Na₂SO₄) and evaporated under reduced pressure. A mixture of the residue in MeOH (5 mL) containing Sm (9.2 mg, 59 μmol) and I₂ (15 mg, 59 μmol) was stirred for 15 min at room temperature. The mixture was evaporated under reduced pressure, and the residue was partitioned between EtOAc (5

mL) and H₂O (2 mL). The organic layer was washed with H₂O (3 mL) and brine (3 mL), dried (Na₂SO₄), and evaporated under reduced pressure. A solution of the residue in THF (5 mL) containing TBAF (1 M THF solution, 50 μ L, 50 μ mol) was stirred for 30 min at 0 °C. The mixture was evaporated under reduced pressure, and the residue was purified by preparative TLC (17% MeOH/CHCl₃) to give herbicidin B (7.3 mg, 31%): $[\alpha]_D^{+55.1}$ (c 0.61, MeOH), lit.^{3b} $[\alpha]_D^{+63}$ (c 1, MeOH); ¹H NMR (CD₃OD, 500 MHz) δ 8.59 (s, 1H), 8.11 (s, 1H), 6.08 (s, 1H), 4.58 (t, 1H, *J* = 8.6 Hz), 4.35 (s, 1H), 4.33 (d, 1H, *J* = 2.3 Hz), 4.27 (s, 1H), 4.25 (d, 1H, *J* = 3.3 Hz), 3.90 (s, 1H), 3.62 (s, 3H), 3.37 (t, 1H, *J* = 9.5 Hz), 3.44 (s, 3H), 2.14 (br s, 1H), 2.12 (s, 1H); ¹³C NMR (CD₃OD, 125 MHz) δ 171.8, 154.0, 142.4, 94.6, 92.2, 89.1, 79.7, 78.0, 74.4, 73.8, 71.3, 65.5, 58.4, 26.4; MS (FAB) *m/z* 454 (MH⁺). Anal. Calcd for C₁₈H₂₃N₅O₉: C, 47.68; H, 5.11; N, 15.45.

Found: C, 47.89; H, 5.09; N, 14.78.

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Supporting Information Available: General experimental methods, experimental details for the synthesis of **9–13**, **19a,b**, and **20–23**, and complete ¹H NMR spectral information for **22**, **23**, **25**, **27b**, and synthetic and natural herbicidin B (PDF). This material is available free of charge via the Internet at <http://pubs.acs.org>.

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